CDK5/p25, Active
Full-length recombinant protein expressed in Sf9 cells

Catalog # C33-10G-10
Lot # P083-1

Product Description
Recombinant full-length human CDK5 and p25 were co-expressed by baculovirus in Sf9 insect cells using an N-terminal GST tag on both proteins. The gene accession numbers for CDK5 and p25 are NM_004935 and NM_003885, respectively.

Gene Aliases
CDK5: PSSALRE
p25: CDK5R1; CDK5P35, CDK5R, NCK5A, p23, p35, p35nc k5a

Concentration
0.1 µg/µl

Formulation
Recombinant protein stored in 50mM Tris-HCl, pH 7.5, 150mM NaCl, 10mM glutathione, 0.1mM EDTA, 0.25mM DTT, 0.1mM PMSF, 25% glycerol.

Storage, Shipping and Stability
Store product at –70°C. For optimal storage, aliquot target into smaller quantities after centrifugation and store at recommended temperature. For most favorable performance, avoid repeated handling and multiple freeze/thaw cycles. Stability is 1yr at –70°C from date of shipment. Product shipped on dry ice.

Scientific Background
CDK5 is a member of the Cyclin-Dependent Kinase family that is most abundant in the mammalian brain. Active form of CDK5, which has also been called neuronal cdc2-like kinase, is a heterodimer of CDK5 and a 25 kDa protein which is derived proteolytically from a 35 kDa brain and neuron-specific protein and is essential for the kinase activity of CDK5 (1). CDK5 has emerged as a crucial regulator of neuronal migration in the developing central nervous system. CDK5 phosphorylates a diverse list of substrates, implicating it in the regulation of a range of cellular processes - from adhesion and motility, to synaptic plasticity and drug addiction (2).

References

Purity
The purity was determined to be >90% by densitometry.
CDK5 Approx. MW 59kDa and p25 Approx. MW 49kDa

Specific Activity
The specific activity of CDK5/p25 was determined to be 686 nmol/min/mg as per activity assay protocol.

FOR IN VITRO RESEARCH PURPOSES ONLY. NOT INTENDED FOR USE IN HUMAN OR ANIMALS.
Activity Assay Protocol

Reaction Components

Active Kinase (Catalog #: C33-10G-10)
Active CDK5/p25 (0.1 μg/μl) diluted with Kinase Dilution Buffer I (Catalog #: K21-09) and assayed as outlined in sample activity plot. (Note: these are suggested working dilutions and it is recommended that the researcher perform a serial dilution of Active CDK5/p25 for optimal results).

Kinase Dilution Buffer I (Catalog #: K21-09)
Kinase Assay Buffer I (Catalog #: K01-09) diluted at a 1:4 ratio (5X dilution) with distilled H₂O.

Kinase Assay Buffer I (Catalog #: K01-09)
Buffer components: 25mM MOPS, pH 7.2, 12.5mM β-glycerol-phosphate, 25mM MgCl₂, 5mM EGTA, 2mM EDTA. Add 0.25mM DTT to Kinase Assay Buffer prior to use.

[33P]-ATP Assay Cocktail
Prepare 250μM [33P]-ATP Assay Cocktail in a designated radioactive working area by adding the following components: 150μl of 10mM ATP Stock Solution (Catalog #: A50-09), 100μl [33P]-ATP (1mCi/100μl), 5.75ml of Kinase Assay Buffer I (Catalog #: K01-09). Store 1ml aliquots at -20°C.

10mM ATP Stock Solution, pH7.2 (Catalog #: A50-09)
Prepare ATP stock solution by dissolving 55mg of ATP in 10ml of Kinase Assay Buffer I (Catalog #: K01-09). Store 200μl aliquots at -20°C.

Substrate (Catalog #: H10-54N)
Histone H1 diluted in distilled H₂O to a final concentration of 1mg/ml.

Assay Protocol

Step 1. Thaw [33P]-ATP Assay Cocktail in shielded container in a designated radioactive working area.
Step 2. Thaw the Active CDK5/p25, Kinase Assay Buffer, Substrate and Kinase Dilution Buffer on ice.
Step 3. In a pre-cooled microfuge tube, add the following reaction components bringing the initial reaction volume up to 20μl:
  Component 1. 10μl of diluted Active CDK5/p25 (Catalog # C33-10G-10)
  Component 2. 5μl of 1mg/ml stock solution of substrate (Catalog #H10-54N)
  Component 3. 5μl distilled H₂O (4°C)
Step 4. Set up the blank control as outlined in step 3, excluding the addition of the substrate. Replace the substrate with an equal volume of distilled H₂O.
Step 5. Initiate the reaction by the addition of 5 μl [33P]-ATP Assay Cocktail bringing the final volume up to 25μl and incubate the mixture in a water bath at 30°C for 15 minutes.
Step 6. After the 15 minute incubation period, terminate the reaction by spotting 20 μl of the reaction mixture onto individual pre-cut strips of phosphocellulose P81 paper.
Step 7. Air dry the pre-cut P81 strip and sequentially wash in a 1% phosphoric acid solution (dilute 10ml of phosphoric acid and make a 1L solution with distilled H₂O) with constant gentle stirring. It is recommended that the strips be washed a total of 3 intervals for approximately 10 minutes each.
Step 8. Count the radioactivity (cpm) on the P81 paper in the presence of scintillation fluid in a scintillation counter.
Step 9. Determine the corrected cpm by removing the blank control value (see Step 4) for each sample and calculate the kinase specific activity as outlined below.

Calculation of [P³³]-ATP Specific Activity (SA) (cpm/pmol)
Specific activity (SA) = cpm for 5 μl [³³P]-ATP / pmoles of ATP (in 5 μl of a 250 μM ATP stock solution, i.e., 1250 pmoles)

Kinase Specific Activity (SA) (pmol/min/μg or nmol/min/mg)
Corrected cpm from reaction / [(SA of [³³P]-ATP in cpm/pmol)*(Reaction time in min)*(Enzyme amount in μg or mg)]*(Reaction Volume) / (Spot Volume)
ADP-Glo™ Activity Assay Protocol

Reaction Components

CDK5/p25 Kinase Enzyme System  
(Promega, Catalog #:V3231)  
CDK5/p25, Active, 10μg (0.1μg/μl)  
Histone H1, 1ml (1mg/ml)  
Reaction Buffer A (5X), 1.5ml  
DTT solution (0.1M), 25μl  

ADP-Glo™ Kinase Assay Kit  
(Promega, Catalog #: V9101)  
Ultra Pure ATP, 10 mM (0.5ml)  
ADP, 10 mM (0.5ml)  
ADP-Glo™ Reagent (5ml)  
Kinase Detection Buffer (10ml)  
Kinase Detection Substrate (Lyophilized)

Reaction Buffer A (5X)
200mM Tris-HCl, pH 7.5, 100mM MgCl₂ and 0.5μg/μl BSA.

Assay Protocol

The CDK5/p25 assay is performed using the CDK5/p25 Kinase Enzyme System (Promega; Catalog #: V3231) and ADP-Glo™ Kinase Assay kit (Promega; Catalog #: V9101). The CDK5/p25 reaction utilizes ATP and generates ADP. Then the ADP-Glo™ Reagent is added to simultaneously terminate the kinase reaction and deplete the remaining ATP. Finally, the Kinase Detection Reagent is added to convert ADP to ATP and the newly synthesized ATP is converted to light using the luciferase/luciferin reaction. For more detailed protocol regarding the ADP-Glo™ Kinase Assay, see the technical Manual #TM313, available at www.promega.com/tbs/tm313/tm313.html.

Step 1. Thaw the ADP-Glo™ Reagents at ambient temperature. Then prepare Kinase Detection Reagent by mixing Kinase Detection Buffer with the Lyophilized Kinase Detection Substrate. Set aside.

Step 2. Thaw the components of CDK5/p25 Enzyme System, ADP and ATP on ice.

Step 3. Prepare 1ml of 2X Buffer by combining 400μl Reaction Buffer A, 1μl DTT and 599μl of dH₂O.

Step 4. Prepare 1ml of 250μM ATP Assay Solution by adding 25μl ATP solution (10mM) to 500μl of 2X Buffer and 475μl of dH₂O.

Step 5. Prepare diluted CDK5/p25 in 1X Buffer (diluted from 2X buffer) as outlined in sample activity plot. (Note: these are suggested working dilutions and it is recommended that the researcher perform a serial dilution of Active CDK5/p25 for optimal results).

Step 6. In a white 96-well plate (Coming Cat # 3912), add the following reaction components bringing the initial reaction volume up to 20μl:

Component 1. 10μl of diluted Active CDK5/p25
Component 2. 5μl of 1mg/ml stock solution of substrate
Component 3. 5μl of 2X Buffer

Step 7. Set up the blank control as outlined in step 6, excluding the addition of the substrate. Replace the substrate with an equal volume of distilled H₂O.

Step 8. At the same time as the CDK5/p25 kinase reaction, set up an ATP to ADP conversion curve at 50μM ATP/ADP range as described in the ADP-Glo™ Kinase Assay technical Manual #TM313.

Step 9. Initiate the CDK5/p25 reactions by the addition of 5μl of 250 μM ATP Assay Solution thereby bringing the final volume up to 25μl. Shake the plate and incubate the reaction mixture at 30°C for 15 minutes.

Step 10. Terminate the reaction and deplete the remaining ATP by adding 25μl of ADP-Glo™ Reagent. Shake the 96-well plate and then incubate the reaction mixture for another 40 minute at ambient temperature.

Step 11. Add 50μl of the Kinase Detection Reagent, shake the plate and then incubate the reaction mixture for another 30 minute at ambient temperature.

Step 12. Read the 96-well reaction plate using the Kinase-Glo™ Luminescence Protocol on a GloMax® Microplate Luminometer (Promega; Cat # E6501).

Step 13. Using the conversion curve, determine the amount of ADP produced (nmol) in the presence (step 6) and absence of substrate (Step 7) and calculate the kinase specific activity as outlined below. For a detailed protocol of how to determine nmols from RLUs, see ADP-Glo™ Applications Database at http://www.promega.com/applications/cellularanalysis/cellsignaling.htm

Kinase Specific Activity (SA) (nmol/min/mg)

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\text{(ADP (step 6) – ADP (Step 7)) in nmol) / (Reaction time in min) * (Enzyme amount in mg)}
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