PIM2, Active
Full-length recombinant protein expressed in Sf9 cells
Catalog # P36-10G-10
Lot # B067-2

Product Description
Recombinant full-length human PIM2 was expressed by baculovirus in Sf9 insect cells using an N-terminal GST tag. The gene accession number is NM_006875.

Gene Aliases
(None)

Concentration
0.1 µg/µl

Formulation
Recombinant protein stored in 50mM Tris-HCl, pH 7.5, 150mM NaCl, 0.25mM DTT, 0.1mM EGTA, 0.1mM EDTA, 0.1mM PMSF, 25% glycerol.

Storage, Shipping and Stability
Store product at –70°C. For optimal storage, aliquot target into smaller quantities after centrifugation and store at recommended temperature. For most favorable performance, avoid repeated handling and multiple freeze/thaw cycles. Stability is 1yr at –70°C from date of shipment. Product shipped on dry ice.

Scientific Background
PIM2 is a serine threonine kinase that is present in all tissues, being most abundant in hematopoietic tissues, spleen, thymus, and peripheral blood leukocytes, as well as in testis, small intestine, and colon (1). It is highly expressed in human leukemic, lymphoma and colorectal adenocarcinoma cell lines. This suggests a role for PIM2 in proliferating cells as well as during meiosis. Similar to PIM1, PIM2 also acts as a pro-survival kinase and BAD protein is a legitimate PIM2 substrate (2).

References

Purity
The purity of PIM2 was determined to be >95% by densitometry. Approx. MW 61kDa.

Figure 1. SDS-PAGE gel image

Figure 2. Radiometric Assay Data
The specific activity of PIM2 was determined to be 305 nmol/min/mg as per activity assay protocol.
(For Radiometric Assay Protocol on this product please see pg. 2)

Figure 3. ADP-Glo™ Assay Data
The specific activity of PIM2 was determined to be 680 nmol/min/mg as per activity assay protocol.
(For ADP-Glo™ Assay Protocol on this product please see pg. 3)
Activity Assay Protocol

Reaction Components

Active Kinase (Catalog #: P36-10G)
Active PIM2 (0.1µg/µl) diluted with Kinase Dilution Buffer III (Catalog #: K23-09) and assayed as outlined in sample activity plot. (Note: these are suggested working dilutions and it is recommended that the researcher perform a serial dilution of Active PIM2 for optimal results).

Kinase Dilution Buffer III (Catalog #: K23-09)
Kinase Assay Buffer I (Catalog #: K01-09) diluted at a 1:4 ratio (5X dilution) with 50 ng/µl BSA solution.

Kinase Assay Buffer I (Catalog #: K01-09)
Buffer components: 25mM MOPS, pH 7.2, 12.5mM β-glycerol-phosphate, 25mM MgCl2, 5mM EGTA, 2mM EDTA. Add 0.25mM DTT to Kinase Assay Buffer prior to use.

Active PIM2 (Catalog #: P36-10G)
[33P]-ATP Assay Cocktail
Prepare 250µM [33P]-ATP Assay Cocktail in a designated radioactive working area by adding the following components: 150µl of 10mM ATP Stock Solution (Catalog #: A50-09), 100µl [33P]-ATP (1mCi/100µl), 5.75ml of Kinase Assay Buffer I (Catalog #: K01-09). Store 1ml aliquots at –20°C.

10mM ATP Stock Solution (Catalog #: A50-09)
Prepare ATP stock solution by dissolving 55mg of ATP in 10ml of Kinase Assay Buffer I (Catalog #: K01-09). Store 200µl aliquots at –20°C.

Substrate (Catalog #: S05-58)
S6K synthetic peptide substrate (KRRRLASLR) diluted in distilled H2O to a final concentration of 1mg/ml.

Assay Protocol

Step 1. Thaw [33P]-ATP Assay Cocktail in shielded container in a designated radioactive working area.
Step 2. Thaw the Active PIM2, Kinase Assay Buffer, Substrate and Kinase Dilution Buffer on ice.
Step 3. In a pre-cooled microfuge tube, add the following reaction components bringing the initial reaction volume up to 20µl:
   Component 1. 10µl of diluted Active PIM2 (Catalog #P36-10G)
   Component 2. 5µl of 1mg/ml stock solution of substrate (Catalog #S05-58)
   Component 3. 5µl distilled H2O (4°C)

Step 4. Set up the blank control as outlined in step 3, excluding the addition of the substrate. Replace the substrate with an equal volume of distilled H2O.
Step 5. Initiate the reaction by the addition of 5µl [33P]-ATP Assay Cocktail bringing the final volume up to 25µl and incubate the mixture in a water bath at 30°C for 15 minutes.
Step 6. After the 15 minute incubation period, terminate the reaction by spotting 20µl of the reaction mixture onto individual pre-cut strips of phosphocellulose P81 paper.
Step 7. Air dry the pre-cut P81 strip and sequentially wash in a 1% phosphoric acid solution (dilute 10ml of phosphoric acid and make a 1L solution with distilled H2O) with constant gentle stirring. It is recommended that the strips be washed a total of 3 intervals for approximately 10 minutes each.
Step 8. Count the radioactivity (cpm) on the P81 paper in the presence of scintillation fluid in a scintillation counter.
Step 9. Determine the corrected cpm by removing the blank control value (see Step 4) for each sample and calculate the kinase specific activity as outlined below.

Calculation of [P33]-ATP Specific Activity (SA) (cpm/pmol)
Specific activity (SA) = cpm for 5µl [33P]-ATP / pmoles of ATP (in 5µl of a 250µM ATP stock solution, i.e., 1250 pmoles)

Kinase Specific Activity (SA) (pmol/min/µg or nmol/min/mg)
Corrected cpm from reaction / [([SA of 33P-ATP in cpm/pmol]*[Reaction time in min]*[Enzyme amount in µg or mg])*(Spot Volume) / (Reaction Volume)]

FOR IN VITRO RESEARCH PURPOSES ONLY. NOT INTENDED FOR USE IN HUMAN OR ANIMALS.
ADP-Glo™ Activity Assay Protocol

PIM2 Kinase Enzyme System
(Promega, Catalog #: V4034)
- PIM2, Active, 10µg (0.1µg/µl)
- S6K Substrate, 1ml (1mg/ml)
- Reaction Buffer A (5X), 1.5ml
- DTT [0.1M], 25µl

ADP-Glo™ Kinase Assay Kit
(Promega, Catalog #: V9101)
- Ultra Pure ATP solution, 10 mM (0.5ml)
- ADP solution, 10 mM (0.5ml)
- ADP-Glo™ Reagent (5ml)
- Kinase Detection Buffer (10ml)
- Kinase Detection Substrate (Lyophilized)

Reaction Buffer A (5X)
200mM Tris-HCl, pH 7.5, 100mM MgCl₂ and 0.5 mg/ml BSA.

Assay Protocol

The PIM2 assay is performed using the PIM2 Kinase Enzyme System (Promega; Catalog #: V4034) and ADP-Glo™ Kinase Assay kit (Promega; Catalog #: V9101). The PIM2 reaction utilizes ATP and generates ADP. Then the ADP-Glo™ Reagent is added to simultaneously terminate the kinase reaction and deplete the remaining ATP. Finally, the Kinase Detection Reagent is added to convert ADP to ATP and the newly synthesized ATP is converted to light using the luciferase/luciferin reaction. For more detailed protocol regarding the ADP-Glo™ Kinase Assay, see the technical Manual #TM313, available at www.promega.com/tbs/tm313/tm313.html.

**Step 1.** Thaw the ADP-Glo™ Reagents at ambient temperature. Then prepare Kinase Detection Reagent by mixing Kinase Detection Buffer with the Lyophilized Kinase Detection Substrate. Set aside.

**Step 2.** Thaw the components of PIM2 Enzyme System, ADP and ATP on ice.

**Step 3.** Prepare 1ml of 2X Buffer by combining 400µl Reaction Buffer A, 1µl DTT and 599µl of dH₂O.

**Step 4.** Prepare 1ml of 250µM ATP Assay Solution by adding 25µl ATP solution (10mM) to 500µl of 2X Buffer and 475µl of dH₂O.

**Step 5.** Prepare diluted PIM2 in 1X Buffer (diluted from 2X buffer) as outlined in sample activity plot. (Note: these are suggested working dilutions and it is recommended that the researcher perform a serial dilution of Active PIM2 for optimal results).

**Step 6.** In a white 96-well plate (Corning Cat # 3912), add the following reaction components bringing the initial reaction volume up to 20µl:

- Component 1: 10µl of diluted Active PIM2
- Component 2: 5µl of 1mg/ml stock solution of substrate
- Component 3: 5µl of 2X Buffer

**Step 7.** Set up the blank control as outlined in step 6, excluding the addition of the substrate. Replace the substrate with an equal volume of distilled H₂O.

**Step 8.** At the same time as the PIM2 kinase reaction, set up an ATP to ADP conversion curve at 50µM ATP/ADP range as described in the ADP-Glo™ Kinase Assay technical Manual #TM313.

**Step 9.** Initiate the PIM2 reactions by the addition of 5µl of 250 µM ATP Assay Solution thereby bringing the final volume up to 25µl. Shake the plate and incubate the reaction mixture at 30°C for 15 minutes.

**Step 10.** Terminate the reaction and deplete the remaining ATP by adding 25µl of ADP-Glo™ Reagent. Shake the 96-well plate and then incubate the reaction mixture for another 40 minute at ambient temperature.

**Step 11.** Add 50µl of the Kinase Detection Reagent, shake the plate and then incubate the reaction mixture for another 30 minute at ambient temperature.

**Step 12.** Read the 96-well reaction plate using the Kinase-Glo™ Luminescence Protocol on a GloMax® plate reader (Promega; Cat# E7031).

**Step 13.** Using the conversion curve, determine the amount of ADP produced (nmol) in the presence (Step 6) and absence of substrate (Step 7) and calculate the kinase specific activity as outlined below. For a detailed protocol of how to determine nmols from RLUs, see Kinase Enzyme Systems Protocol at: http://www.promega.com/KESProtocol

**Kinase Specific Activity (SA) (nmol/min/mg)**

\[(\text{ADP (step 6) } - \text{ADP (Step 7)}) \text{ in nmol} / (\text{Reaction time in min}) \times (\text{Enzyme amount in mg})\]

For in Vitro Research Purposes Only. Not Intended for Use in Human or Animals.