

A Real-Time *In Vitro* Safety Assessment Approach Utilizing a Simplified, Multi-Parametric Work Flow

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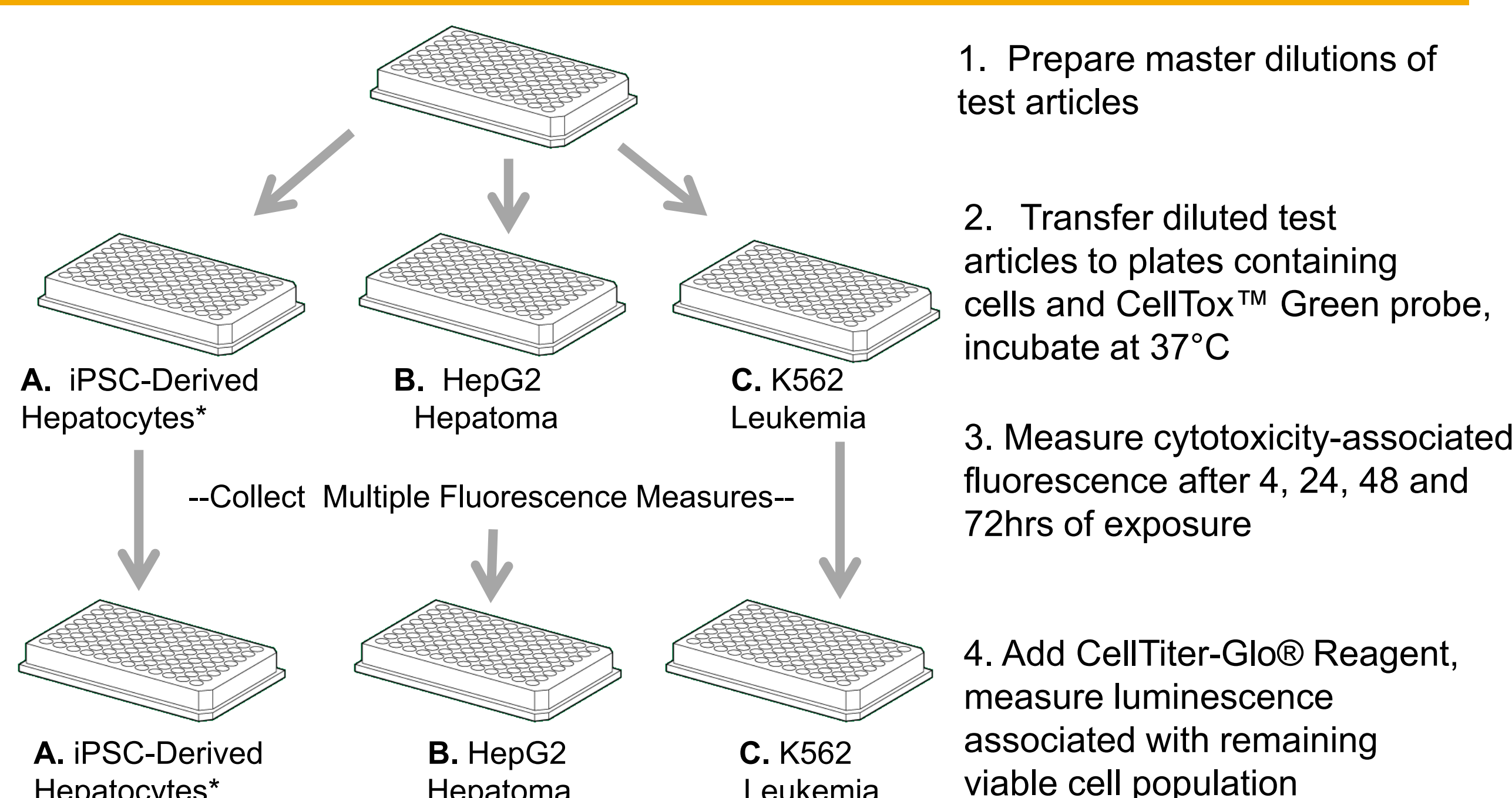
Abstract # 1628



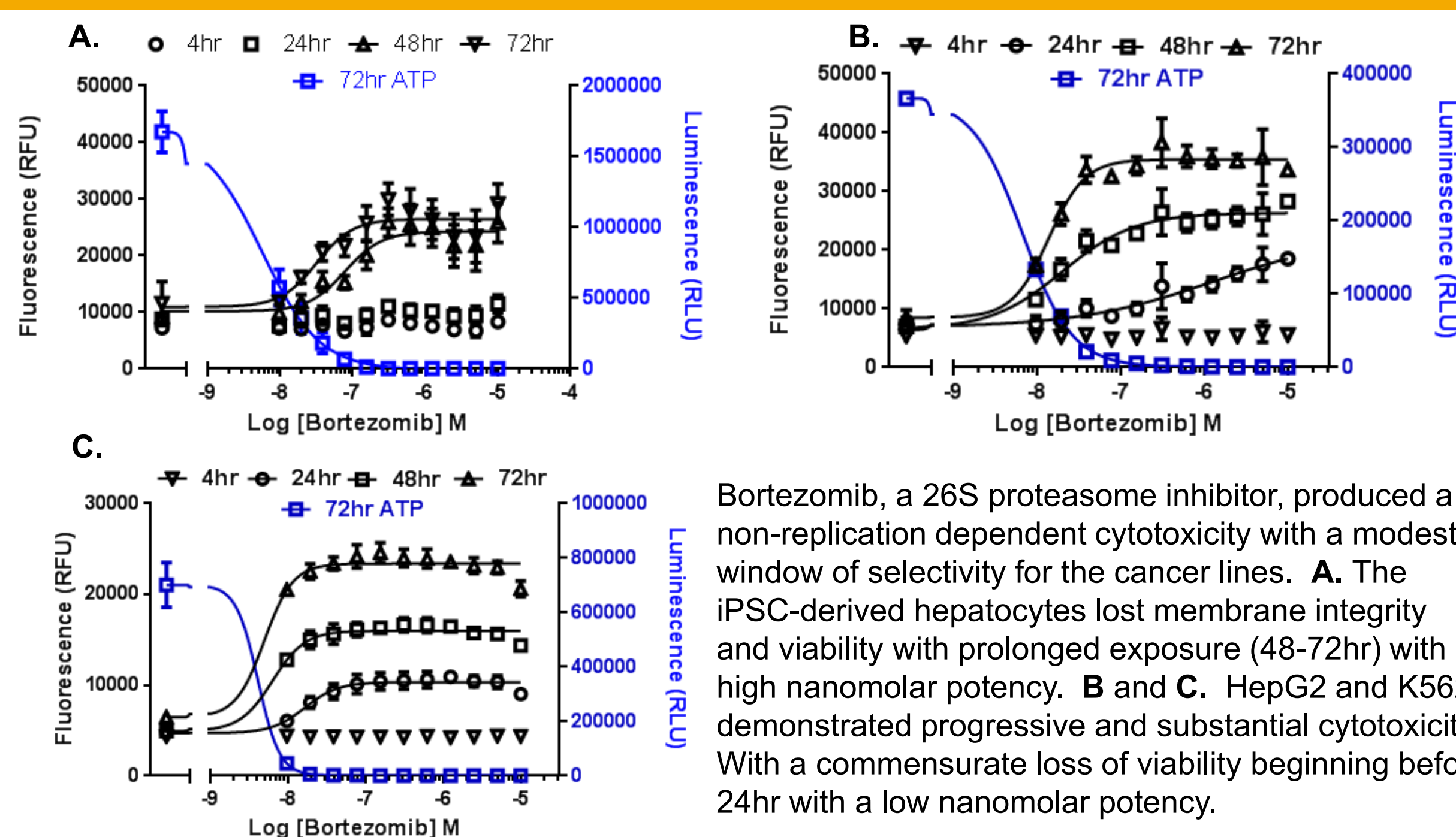
1. Abstract

In vitro cytotoxicity is inextricably linked to a combination of compound dosage, exposure period, and intrinsic cell susceptibility. Current screening paradigms which utilize only endpoint measures in a defined cell type adequately address effects due to dosage, but often fail to define important toxicokinetic profiles or inherent mechanistic sensitivities. We investigated the use of a real-time cytotoxicity probe applied at the time of dosing with staurosporine, panobinostat, imatinib, terfenadine, colchicine, aflatoxin B1, bortezomib, camptothecin, valinomycin, nocodazole, methotrexate and ionomycin. Serial dilutions of these model compounds with the probe were delivered to iPSC-derived, terminally differentiated hepatocytes and proliferating hepatoma and erythroid leukemia cell lines. Cytotoxicity data were collected at 4, 24, 48 and 72hr followed by a same-well multiplexed viability assay. The collated data revealed striking differences in toxicokinetics, potency and magnitude of response which positively correlated with known mechanism of action for the model compounds. The multiplexed viability data further served to either confirm observed cytotoxicity by inverse signal concordance, or suggest replicative perturbation in susceptible replicating cells. Furthermore, the use of cell types with differential capacity for phase I metabolism, allowed us to stratify cytotoxic risk based on mode-of-action of parent molecule toxicity and/or metabolic by-products owing to biotransformation. Lastly, the experimental approach taken was sufficiently predictive and informative to merit consideration for adoption as a new safety screening paradigm for new chemical entities.

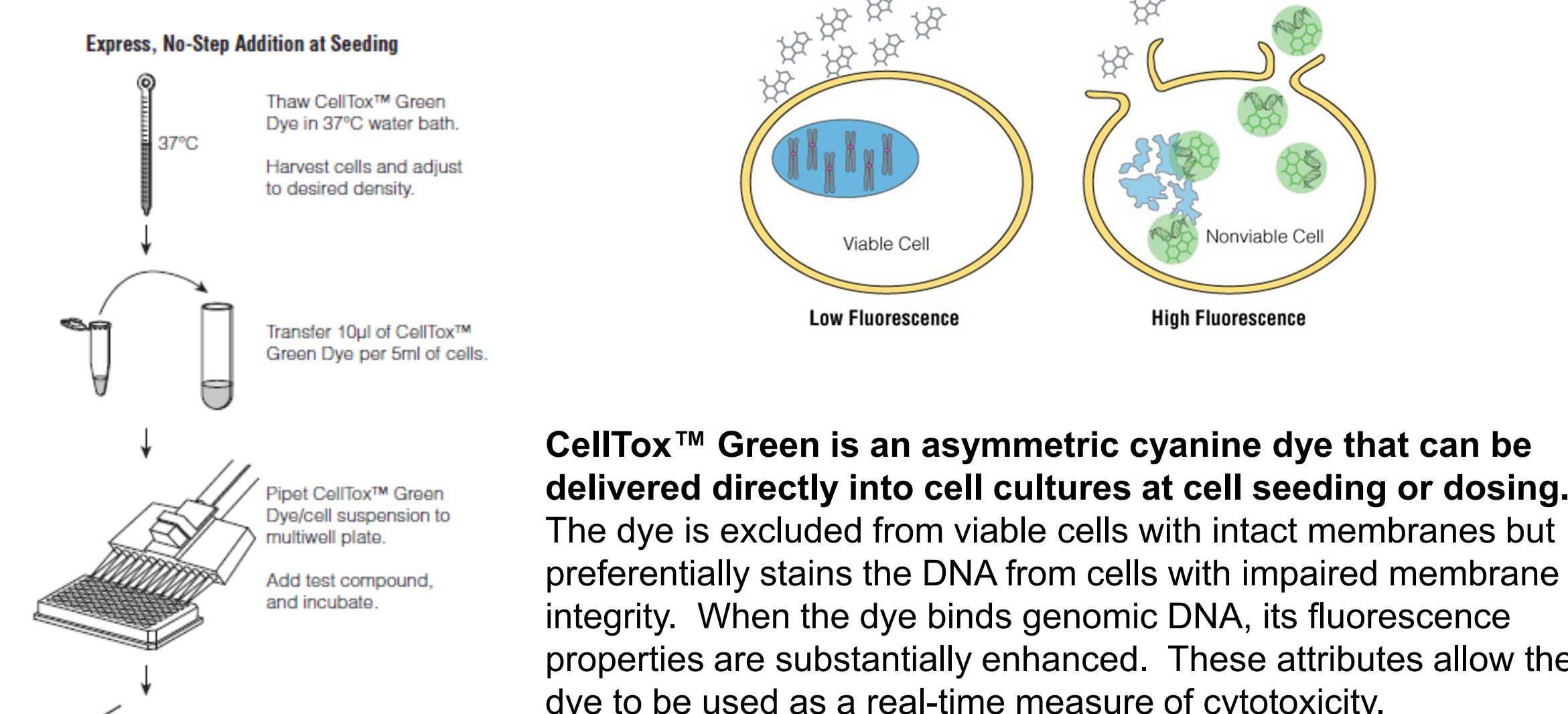
4. Multi-Parametric Workflow: Biomarkers and Cell Types



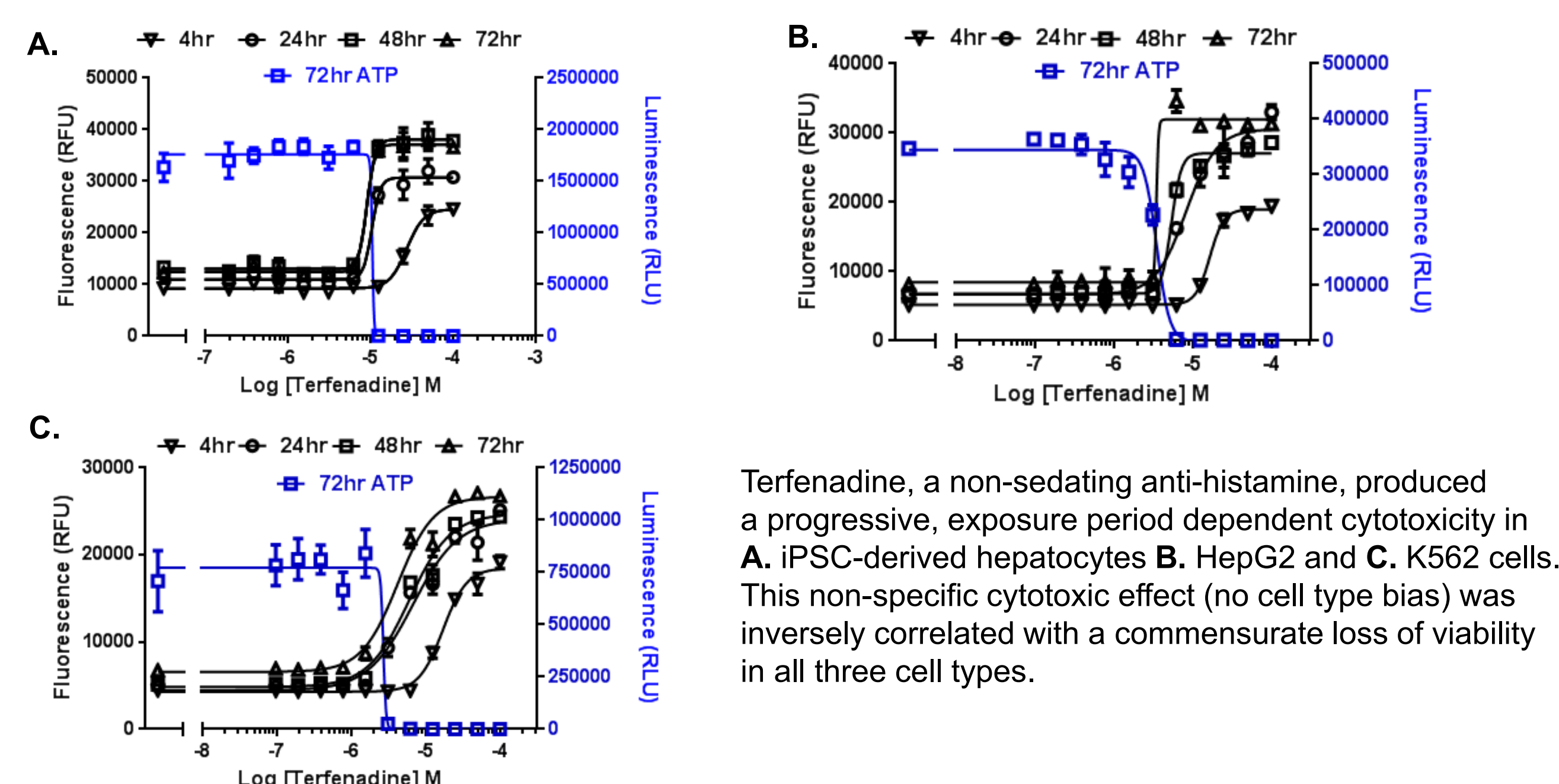
7. Non-Replication Dependent, Targeted Cytotoxicity



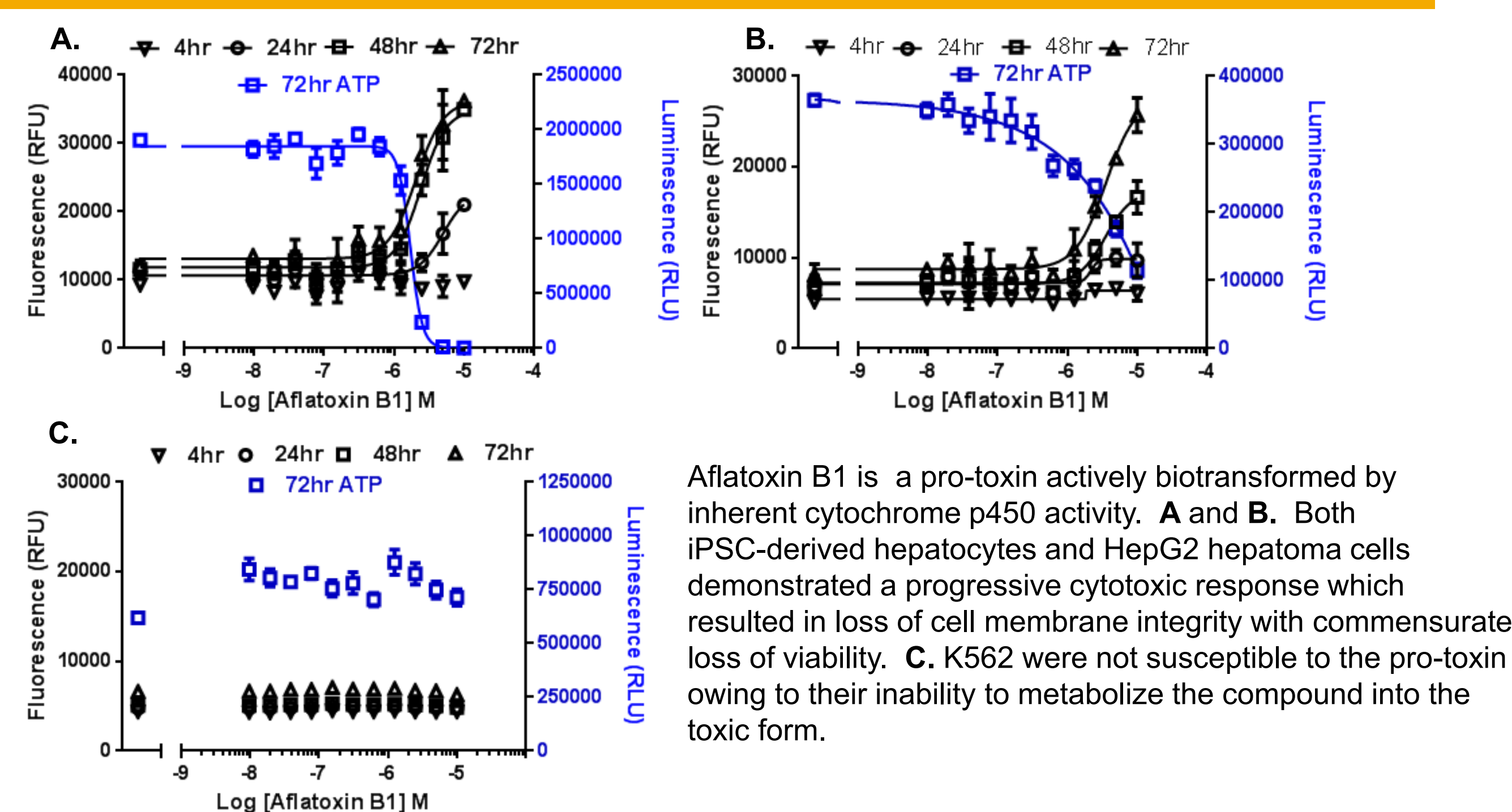
2. Real-Time Cytotoxicity Assay Method and Principle



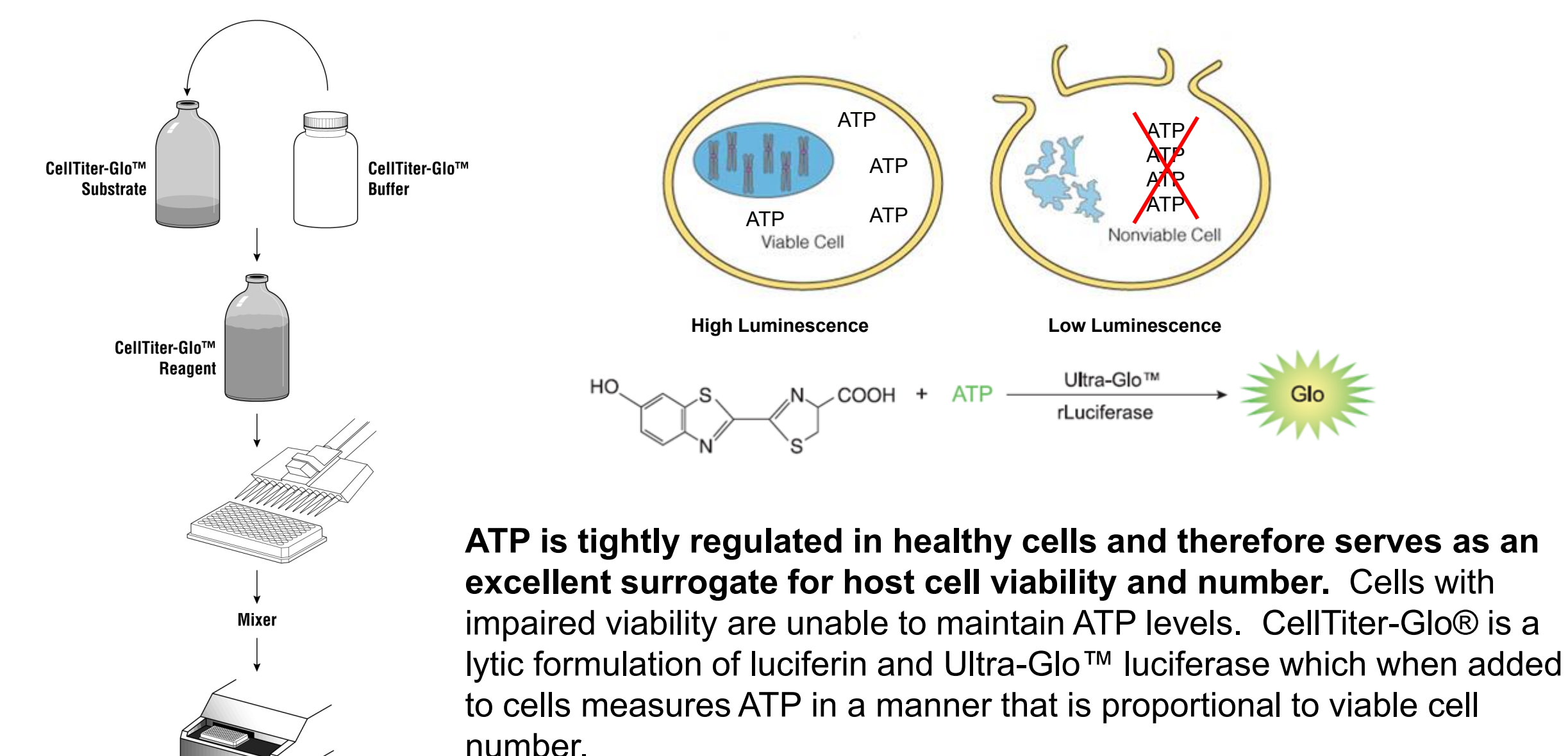
5. Non-Specific, Time Dependent Cytotoxicity



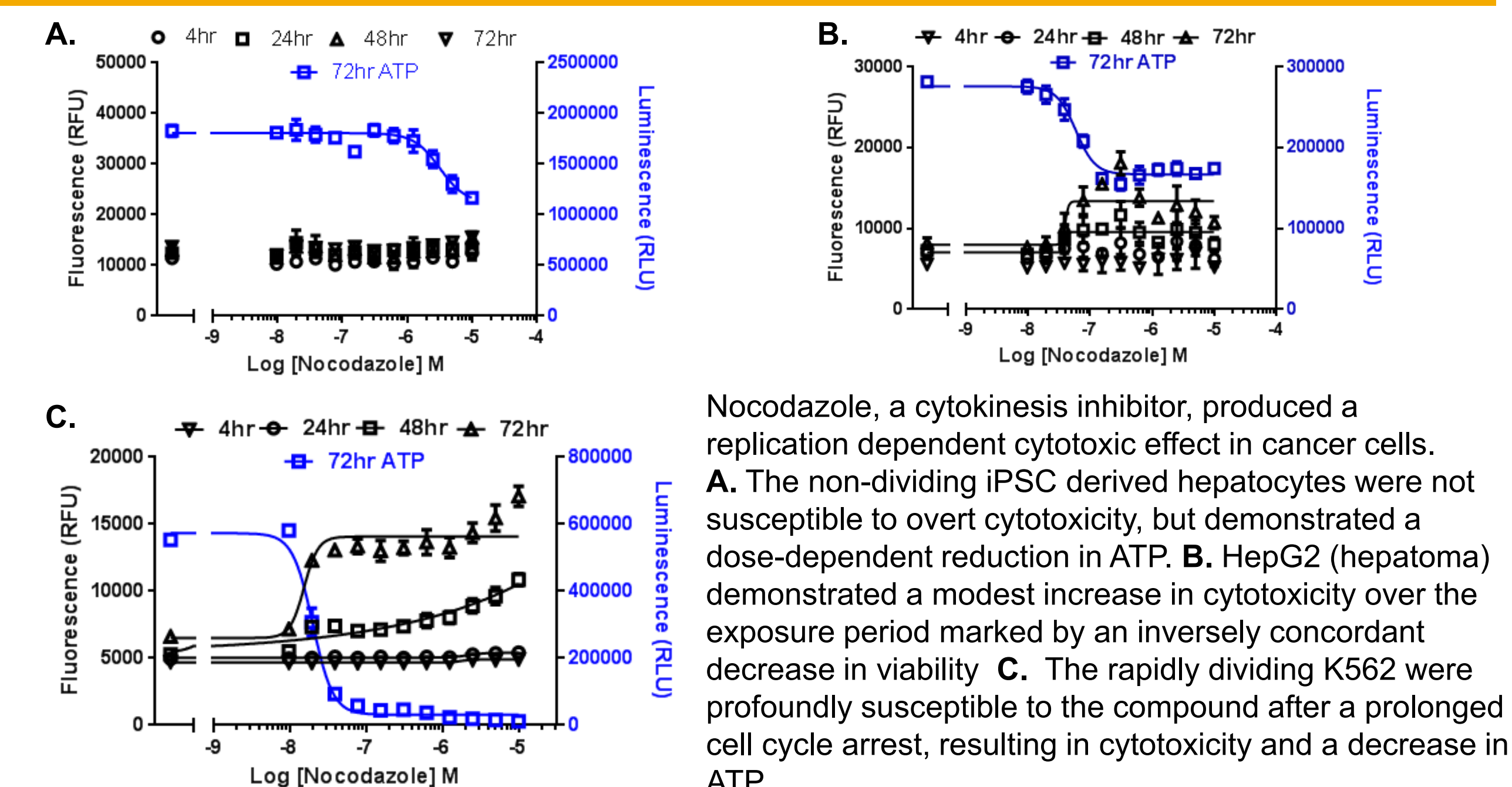
8. Biotransformation Dependent Cytotoxicity



3. Viability Assay Method and Principle



6. Replication Dependent Cytotoxicity



9. Summary

- Introduction of the pro-fluorescent cytotoxicity probe at the time of cell seeding or dosing, allows for a facile and flexible means to measure real-time cytotoxicity. Measurement of cytotoxicity in real-time allows for the development of revealing toxicokinetic profiles for new chemical entities or other test articles.
- Application of the ATP viability chemistry at the terminal endpoint allows for an orthogonal measure of cell health in non-replicating cells, and a measure of overall cell number after xenobiotic exposure.
- Multi-parametric analysis using disparate test cell phenotypes can define mechanism of action for:
 - Non-specific cytotoxic compounds
 - Replication dependent cytotoxicity
 - Targeted anti-neoplastics with on- and off-target efficacy
 - Biotransformed compounds which produce reactive metabolite

* iCell® Hepatocytes were graciously provided through a collaboration with Cellular Dynamics International 525 Science Drive, Madison, WI 53711

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