

# ***Cell-Based Assays to Detect Mechanisms of Toxicity***

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# Overview

## Real-time assays for multi-well plates:

- cell viability
- cell death

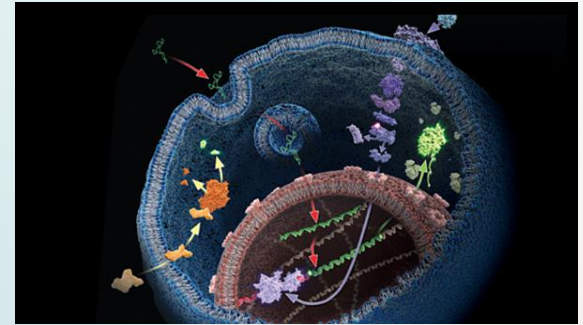
## Cytotoxicity assays:

- biomarkers of apoptosis, necrosis or cell stress

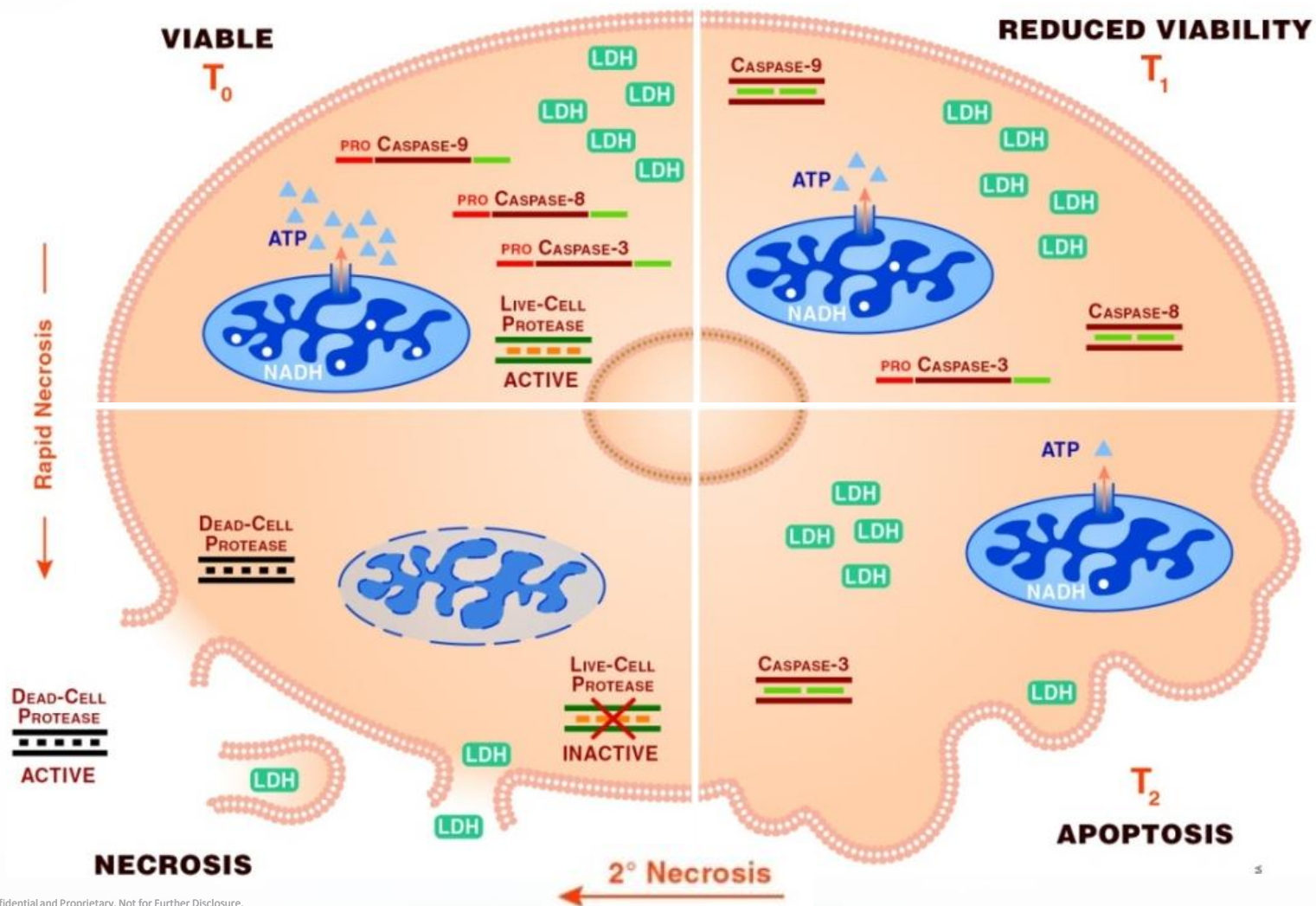
## Multiplexing:

- advantages and examples

# ***Introduction***

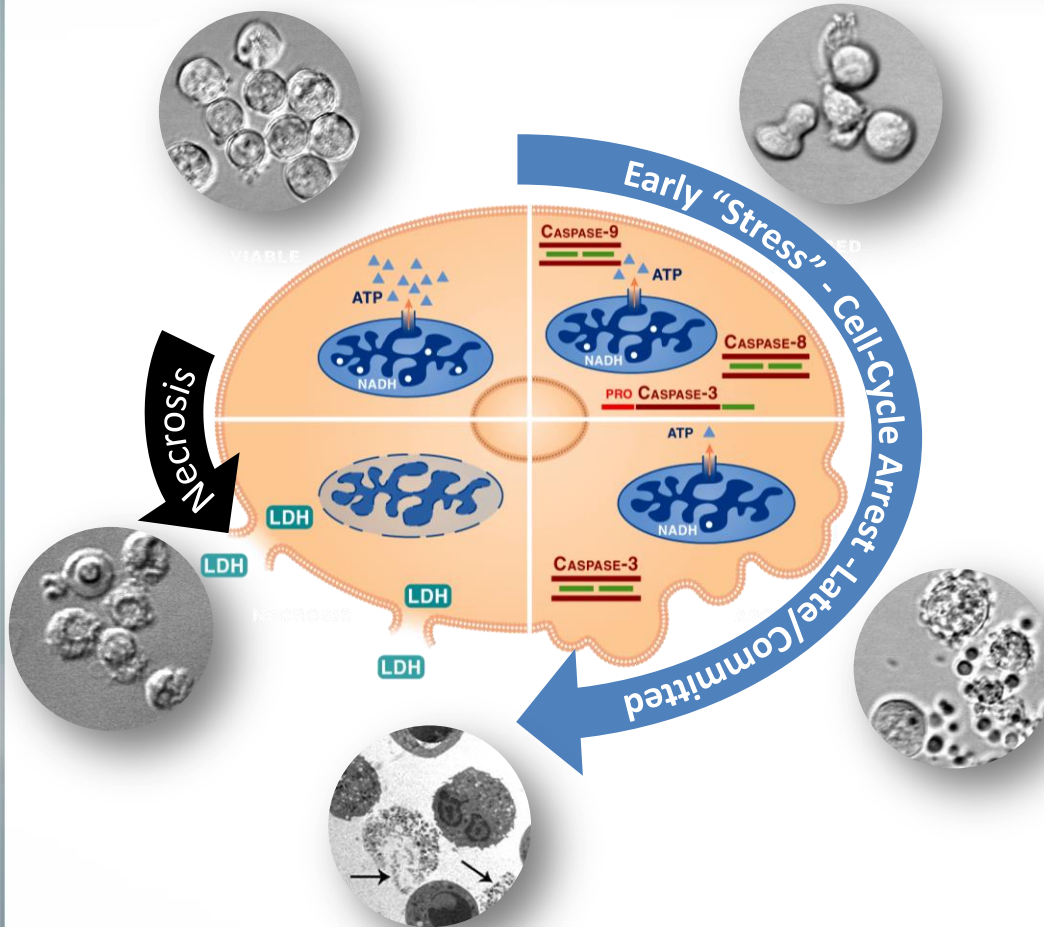


# Monitoring Cytotoxicity





# Cytotoxicity and Biological Complexity



Did the treatment affect viability:

- yes/ no?
- how?
- When?

How **potent** was the treatment?

**Cytotoxicity depends on:**

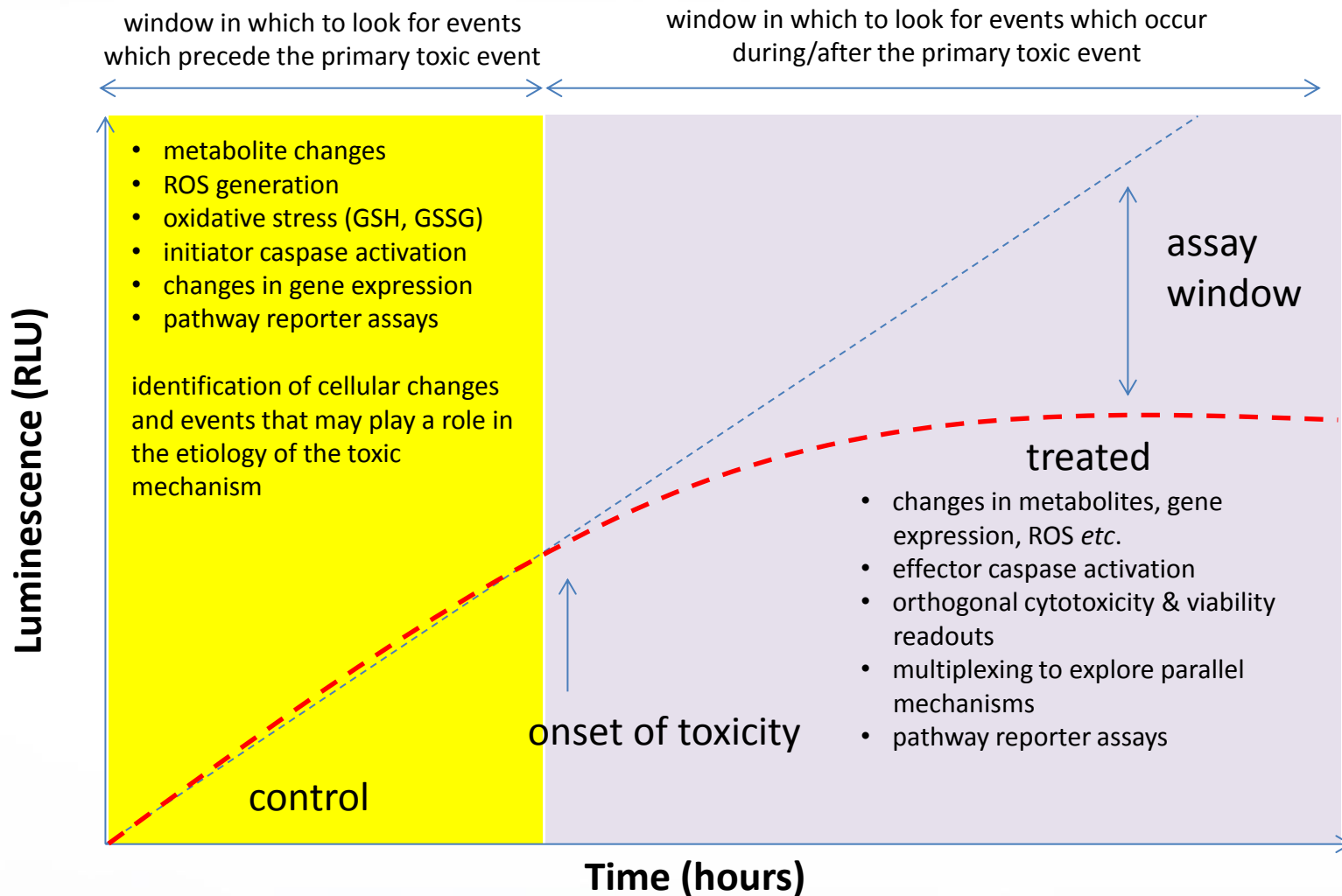
- Dosage
- Exposure time
- Mechanism of action
- Cell type and susceptibility

# ***Cytotoxicity Determination***

<b>Cellular response to treatment</b>	<b>Viability</b>	<b>Cytotoxicity</b>	<b>Apoptosis</b>
<b>Proliferation</b>	↑	→	→
<b>Cell cycle arrest/ senescence</b>	→	→	→
<b>Cytotoxicity (necrosis)</b>	↓	↑	→
<b>Cytotoxicity (apoptosis)</b>	↓	↑	↑

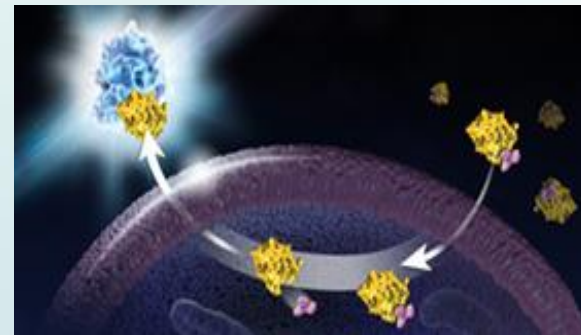
**No single parameter can fully characterise cytotoxicity**

# Timing of Cytotoxicity is Critical!

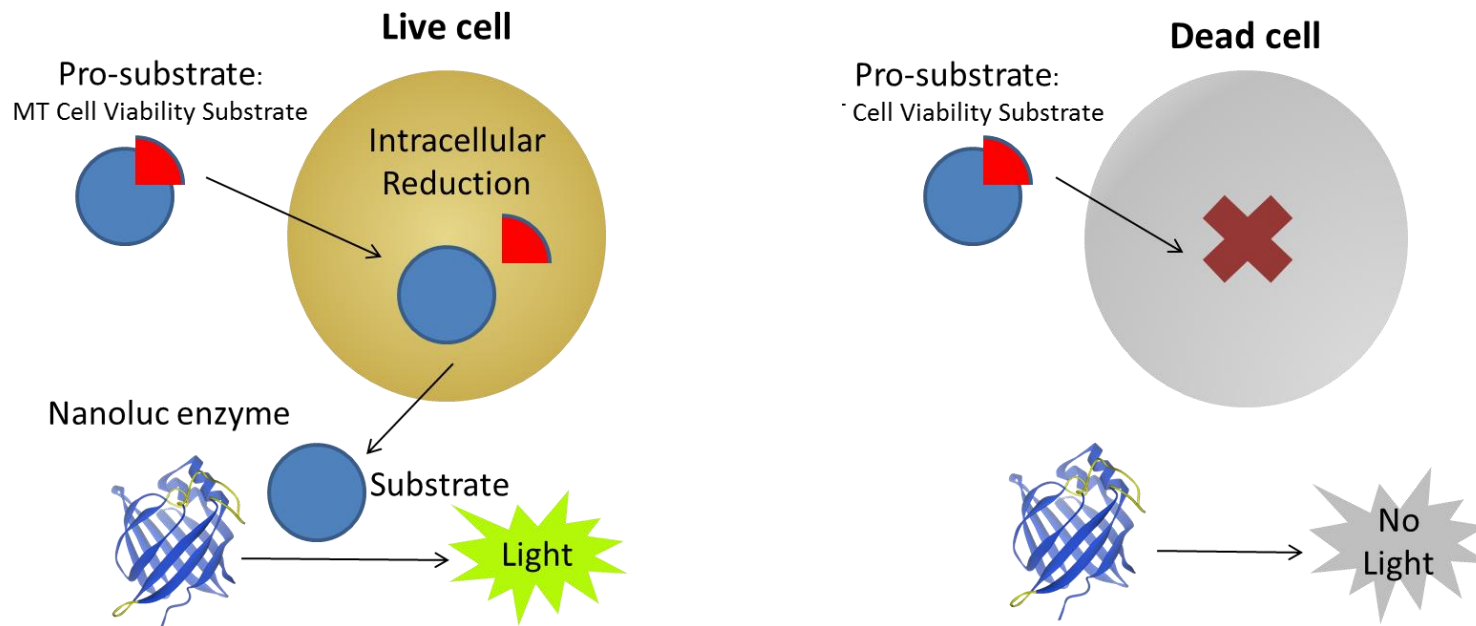


***Real-Time Assay:***

***RealTime-Glo™ MT Cell  
Viability Assay***



# RealTime-Glo™ MT Cell Viability Assay Measures Reducing Potential of Cells

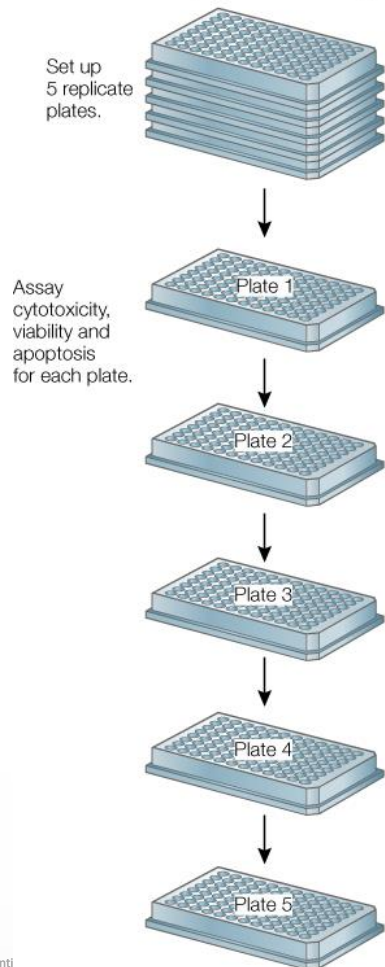


- NanoLuc® enzyme is present outside of the cells
- Pro-substrate enters the cell and is reduced by the cell to form the NanoLuc® substrate
- The NanoLuc® substrate diffuses from the cell and is rapidly used by NanoLuc® to produce light

# Why is RealTime-Glo™ MT Cell Viability Assay Different?

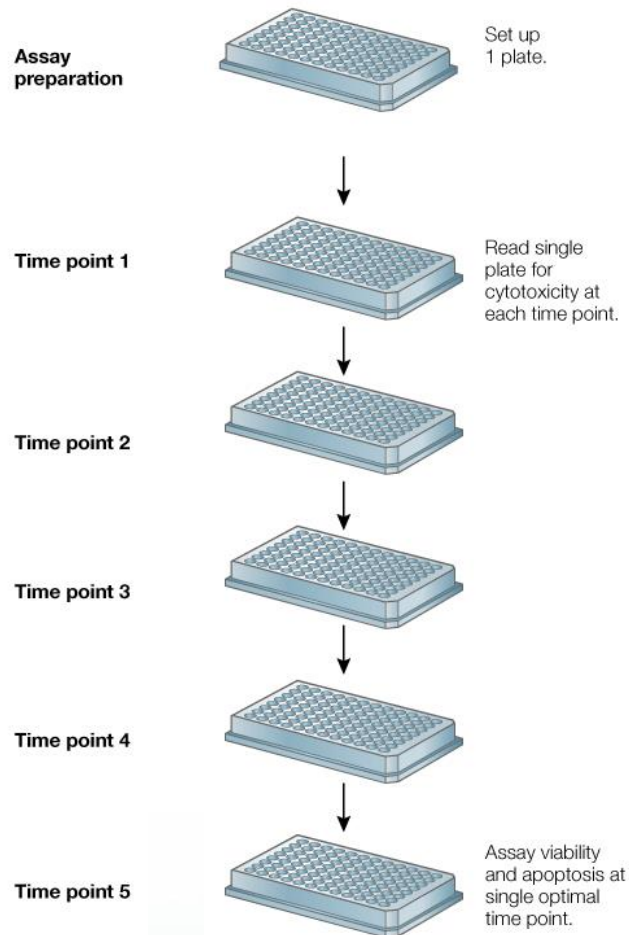
## Traditional approach

### Multiple Time Point Assay



## RealTime-Glo™ MT Cell Viability Assay

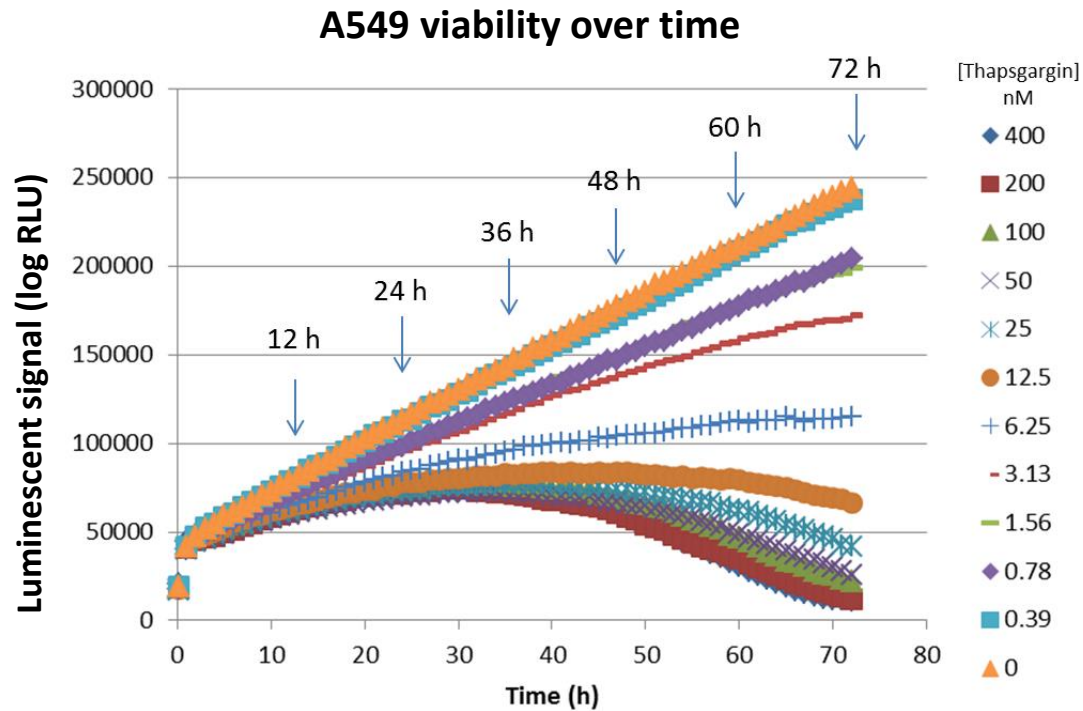
### Kinetic Cytotoxicity Assay



### Advantages:

- Multiple measurements
- Reduced variation & data points
- Cost-effective
- Multiplexing

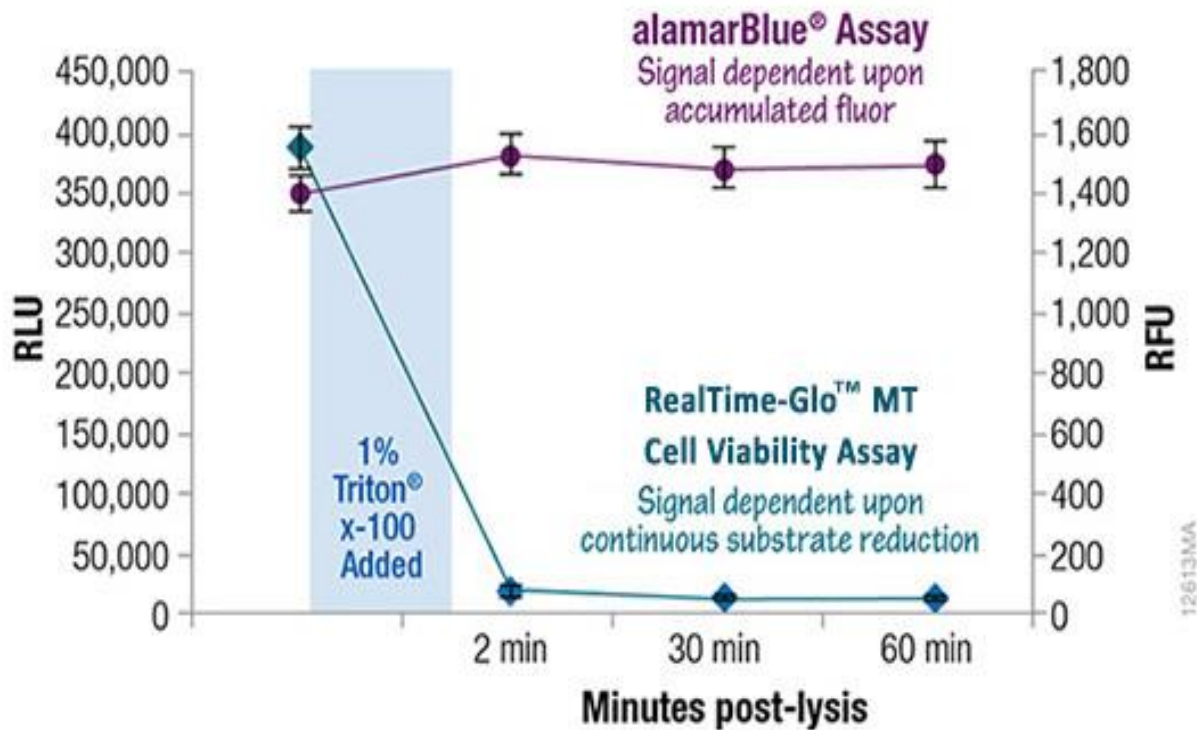
# RealTime-Glo™ Measures Signal in Real-time



A549 cells were treated with thapsigargin dose curve. Luminescence measured continuously every 1h over 72h in Tecan M200 plate reader, with gas control module (37°C/5% CO<sub>2</sub>).

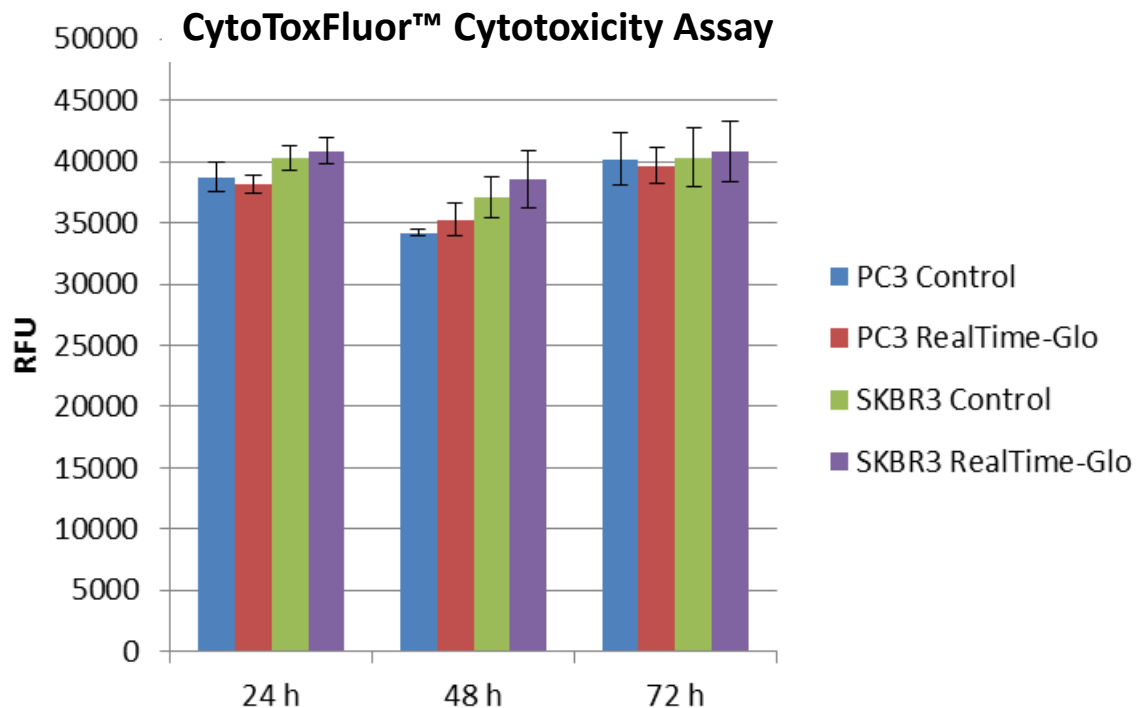
# RealTime-Glo™ is More Responsive

## RealTime-Glo™ vs. AlamarBlue



A549 cells treated with 1% Triton® x-100. Monitored for 1h post- treatment.

## *RealTime-Glo™ Reagents are Not Toxic*

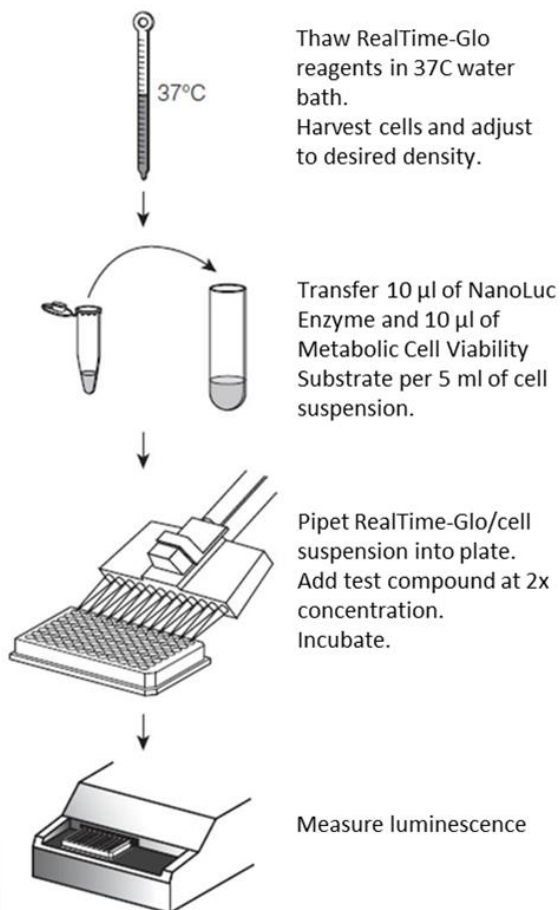


PC3 and SKBR3 cells were incubated in the presence or absence of RealTime-Glo™ reagents for 72h. Samples were tested for membrane integrity using **CytoTox-Fluor™ Cytotoxicity Assay**.

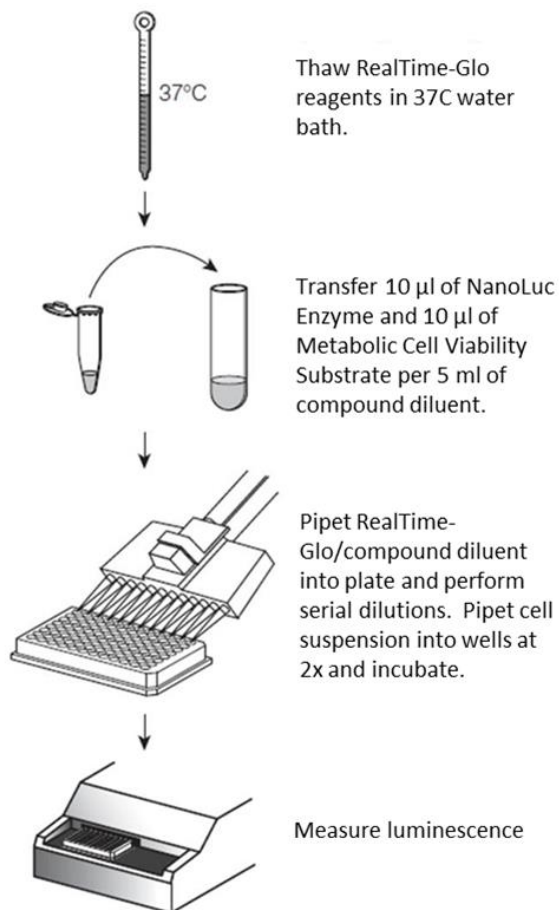
# Different Assay Set up Methods

## RealTime-Glo™ is flexible

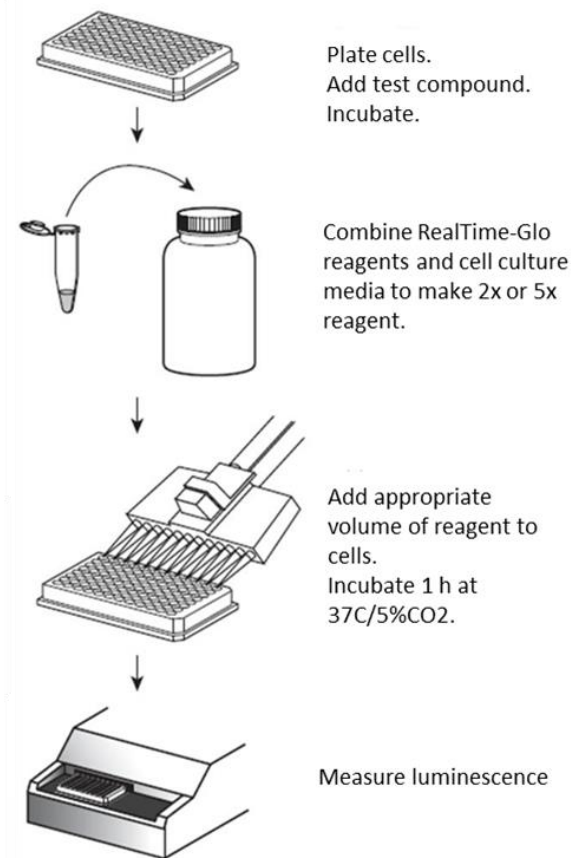
### Continuous Read Format, Addition at Seeding



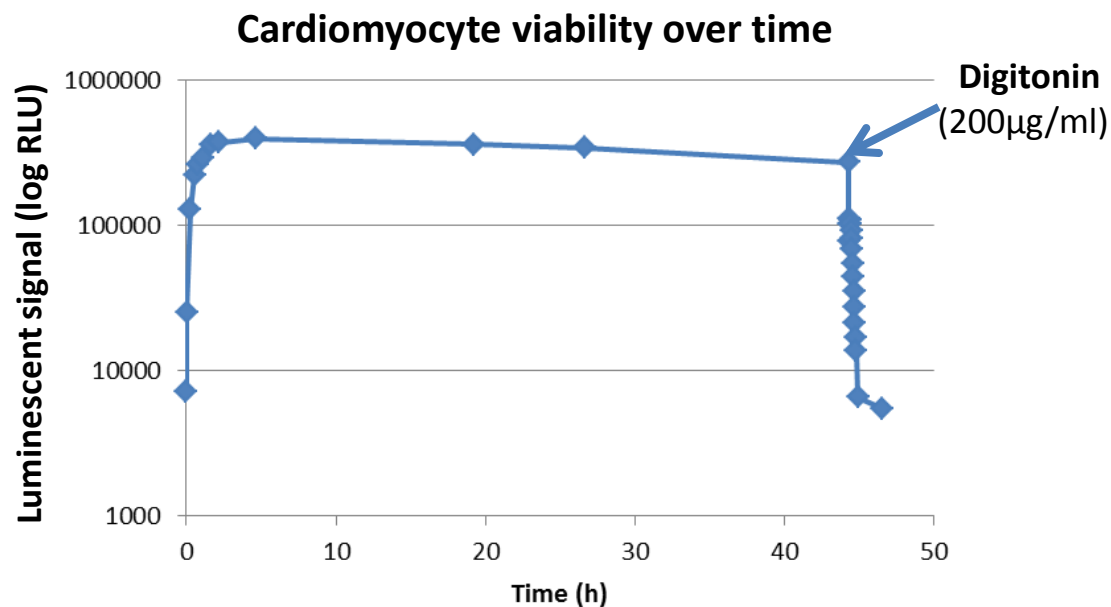
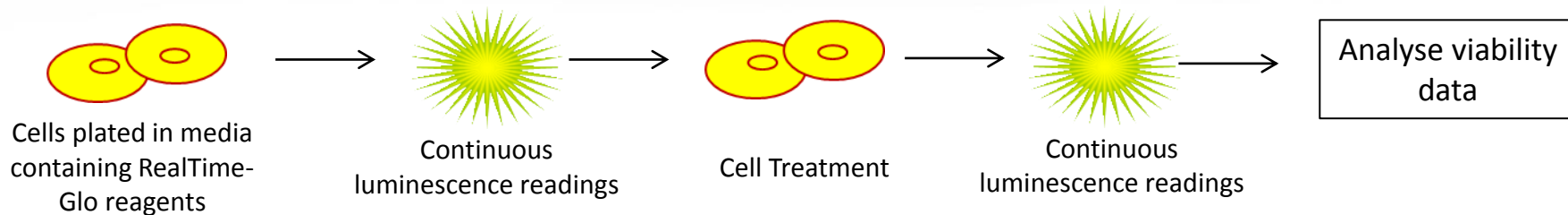
### Continuous Read Format, Addition at Dosing



### Endpoint Method, 2X or 5X Reagent Addition



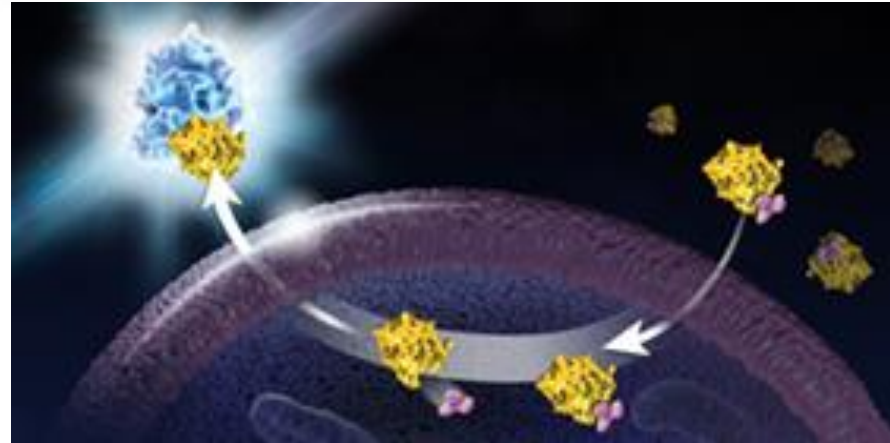
# Continuous Read Format and Responsiveness



## RealTime-Glo™ Summary

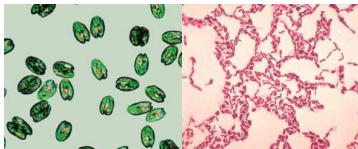
### Features:

- **Real-time** measurements
- Superior **responsiveness**
- **Non- toxic**
- Cost-effective
- Reduces variation & data points
- **Multiplex** opportunities



### Applications:

- Mammalian cells
- Bacteria (gram+ and gram-)
- Yeast and fungus
- Algae

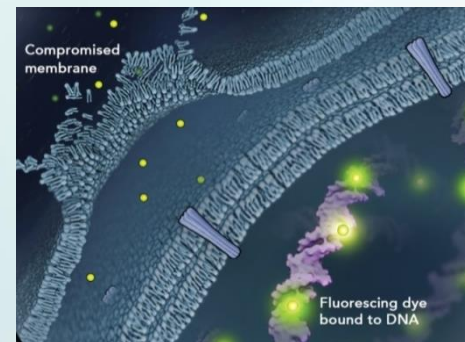


### Kit components:

- NanoLuc® enzyme
- MT Cell Viability Substrate

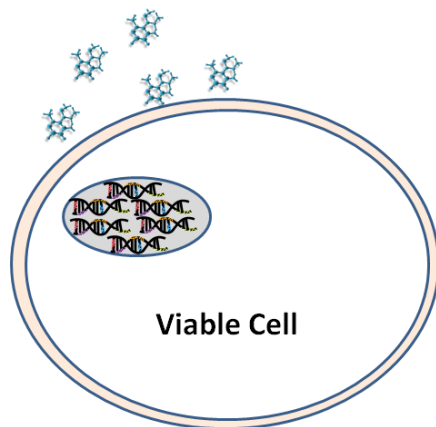
***Real- Time Assay:***

***CellTox™ Green  
Cytotoxicity Assay***

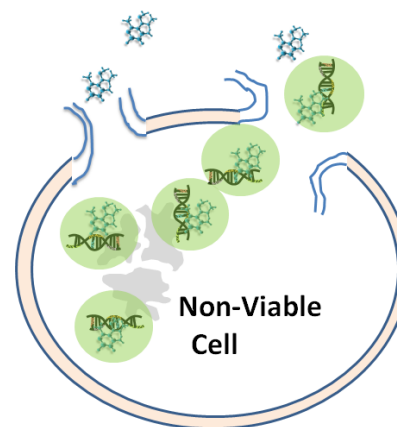


# CellTox™ Green Cytotoxicity Assay

## CellTox™ Green Dye



excluded dye =  
low fluorescence



non-excluded dye =  
high fluorescence

CellTox™ Green Dye spectra

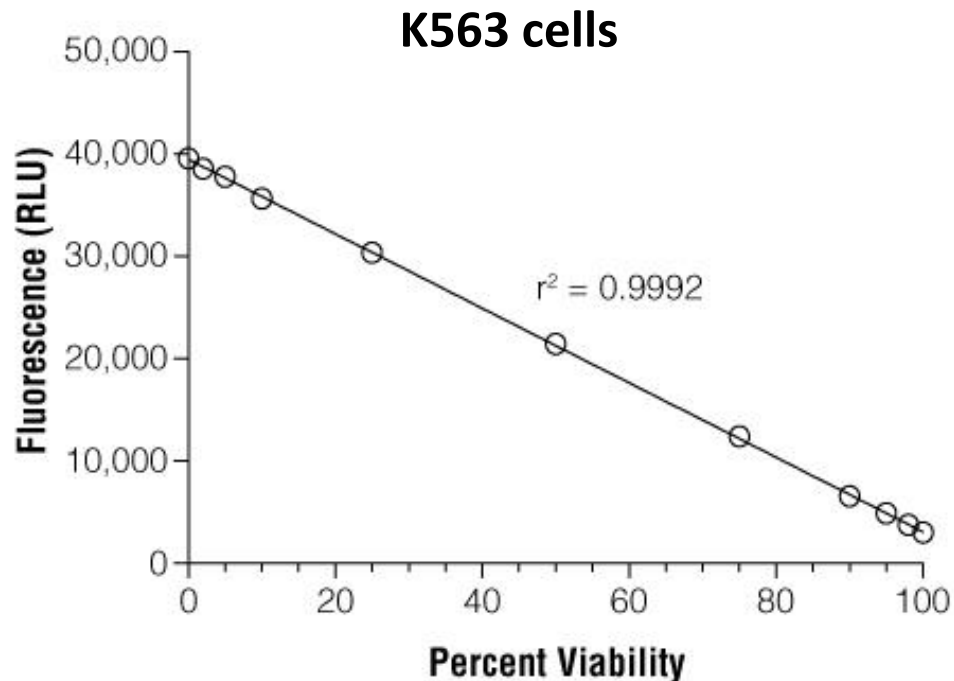
Ex: 512nm

Em: 523nm

Detection: GloMax Multimode 'blue filter'



## ***CellTox™ Green Signal is Directly Proportional to Number of Dead Cells***

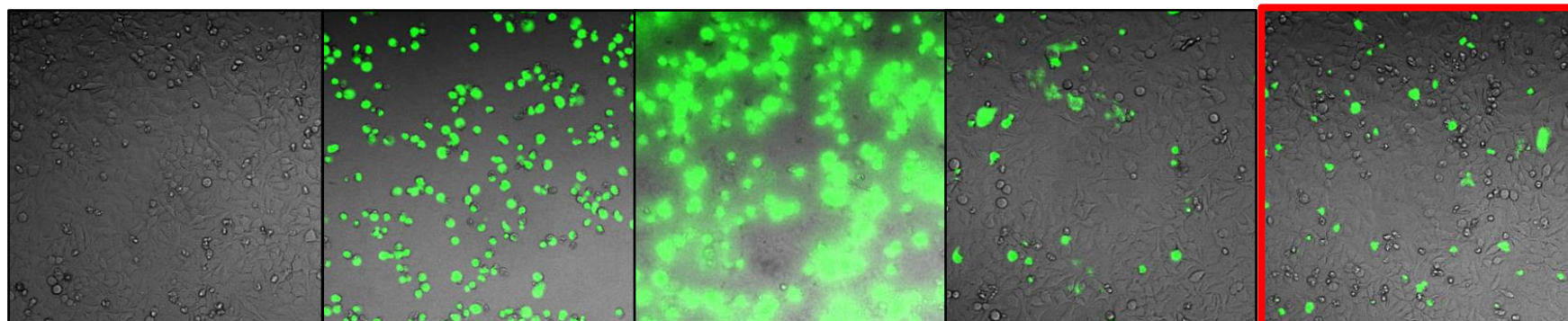


To create a range of prototypical cytotoxic response a population of non-viable K562 cells was mixed with a population of viable K56 cells at various ratios to represent 0- 100% viability. Each ratio contained a total of 10K cells/ 100 $\mu$ l/ well in 96- well plate.

# CellTox™ Green is Not Toxic

Fluorescence (imaging)

HeLa Cells (72h incubation)



Control  
(DMSO)

CyQuant Direct®  
Life Technologies

CyQuant®NF  
Life Technologies

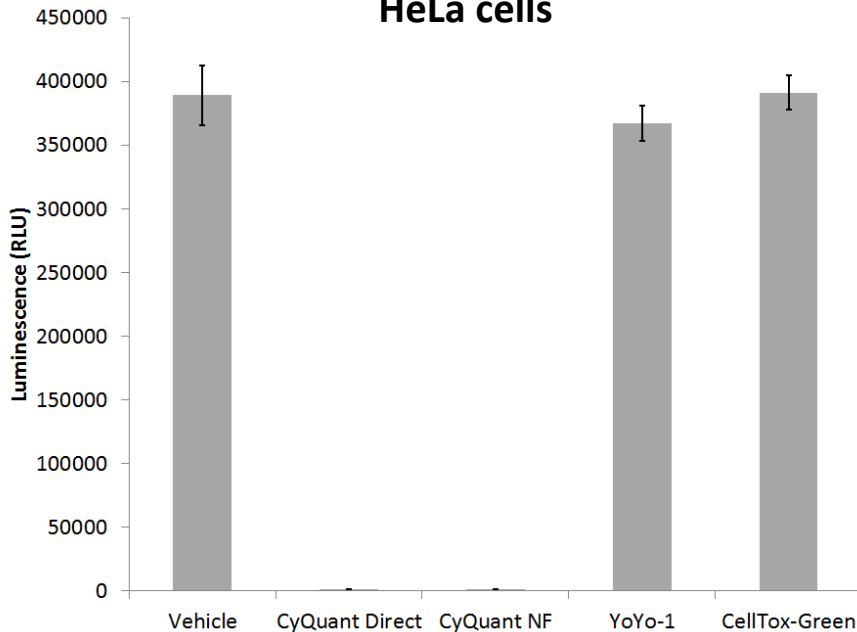
YoYo®-1  
Life Technologies

CellTox™ Green

# CellTox™ Green is Not Toxic

## Luminescence : CellTiter-Glo® Viability Assay

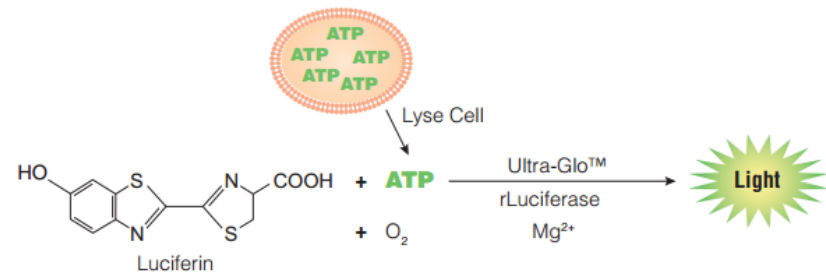
CellTiter Glo® Viability Assay  
HeLa cells



### CellTiter-Glo®

- luminescent assay
- Lytic
- measures ATP
- 'Add-mix- measure'

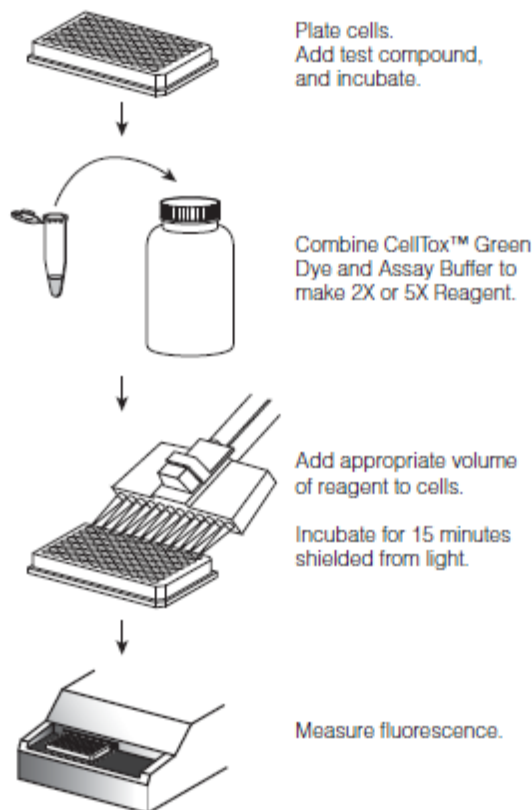
### CellTiter Glo®



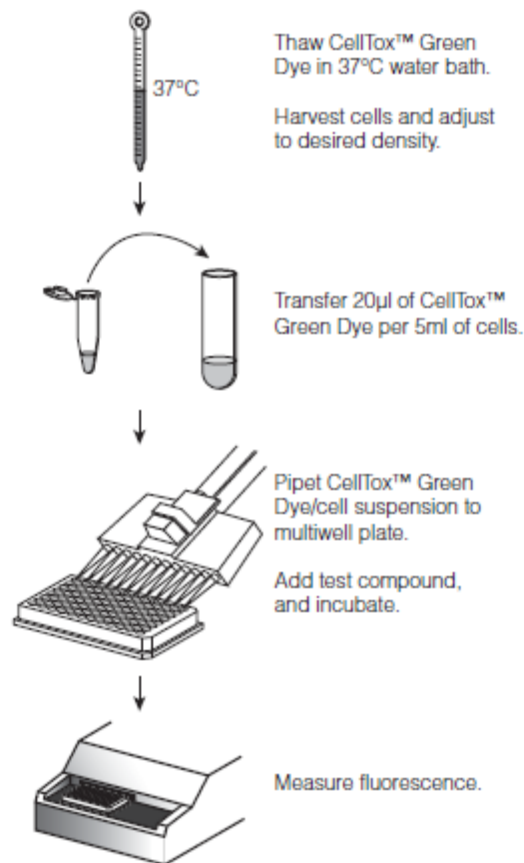
# Different Assay Set up Methods

## CellTox Green™ is flexible

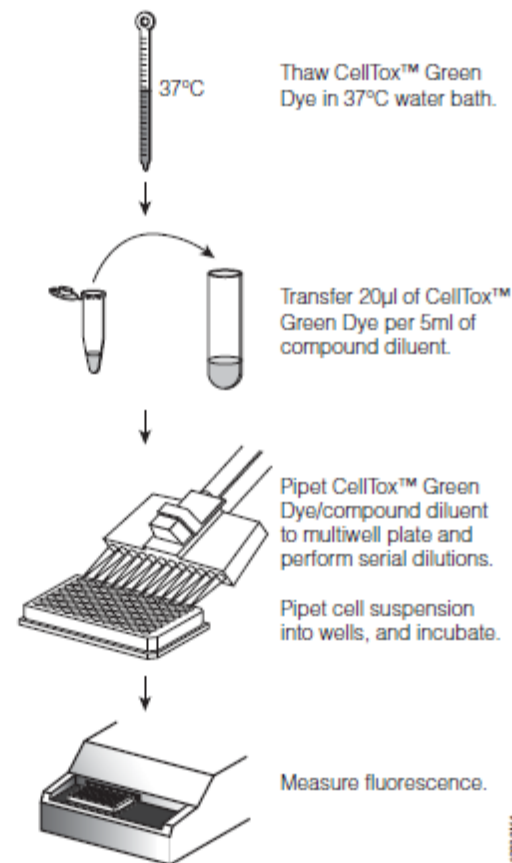
### A. Endpoint Method, 2X or 5X Reagent Addition



### B. Express, No-Step Addition at Seeding



### C. Express, No-Step Addition at Dosing

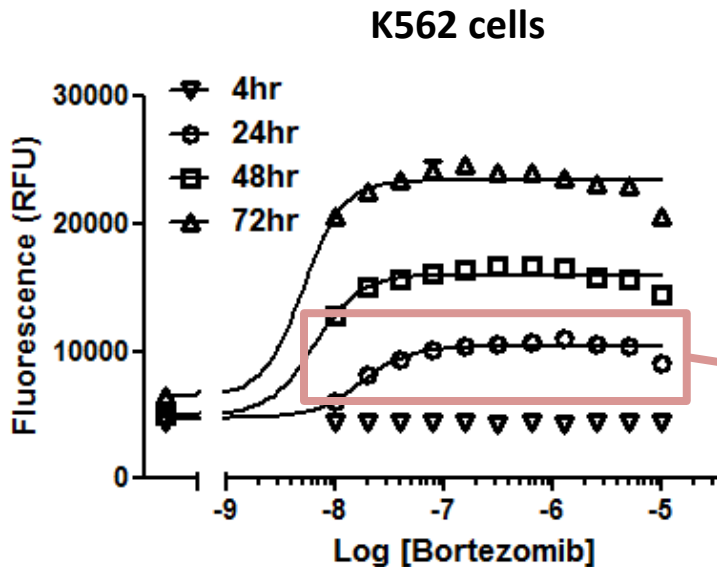


# CellTox™ Green Can Tell You When to Measure Caspase Activity!

CellTox™ Green Cytotoxicity Assay

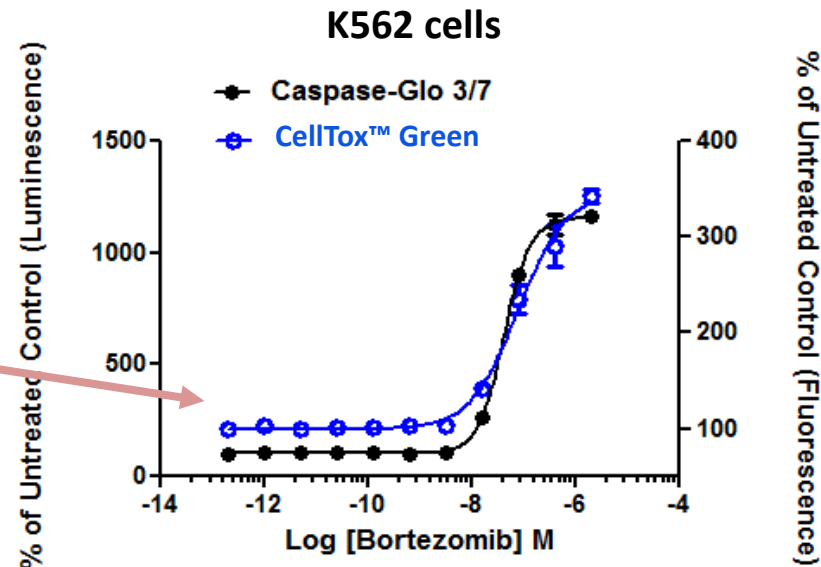


Caspase-Glo® 3/7 Assay



add CellTox™ Green at dosing and monitor for cytotoxicity

K562 cells treated with Bortezomibid for 72h.



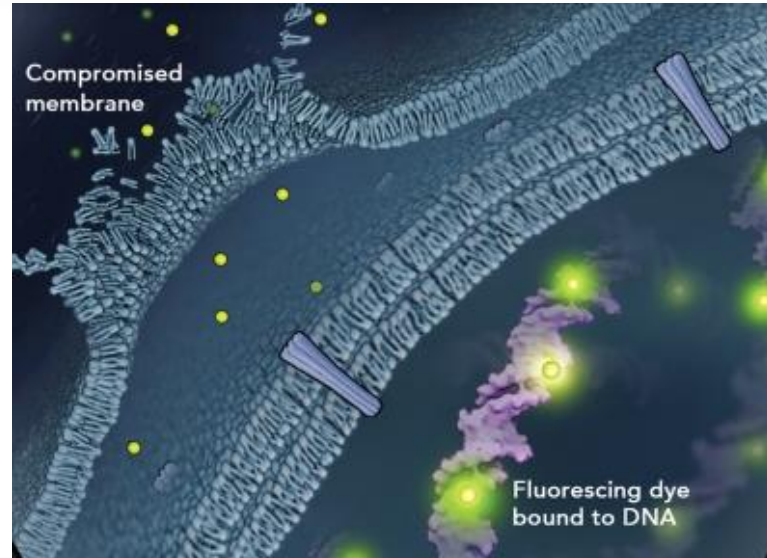
at first emergence of cytotoxicity, add Caspase-Glo® 3/7 Assay reagent

K562 cells treated with Bortezomibid for 24h. CellTox Green and Caspase-Glo 3/7 assays.

# CellTox-Green™ Summary

## Features:

- **Real-time** cell death assay
- **Non-toxic**
- No enzyme involved → no half-life issues
- Cost-effective
- Reduces variation & data points
- **Multiplex** opportunities



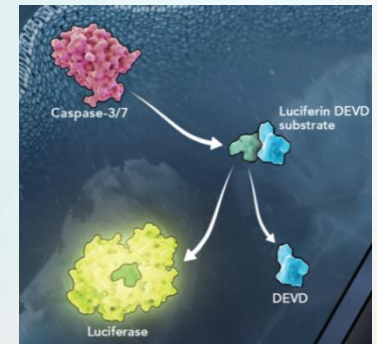
## Applications:

- Mammalian cells

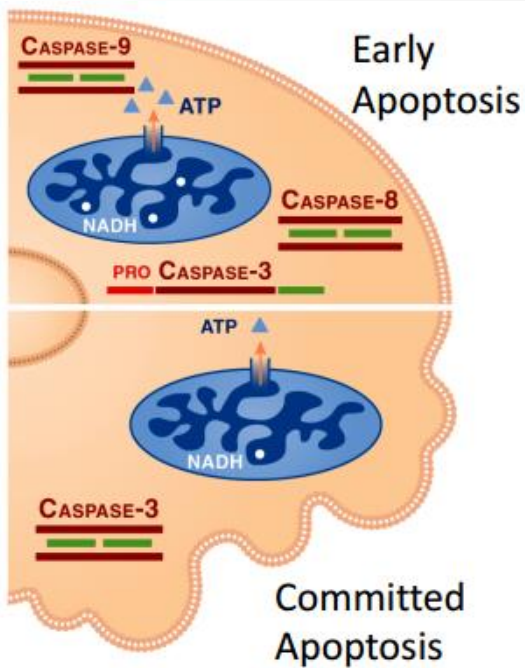
## Kit components:

- CellTox™ Green Dye
- Assay Buffer
- Lysis Buffer

# ***Cytotoxicity Detection: Apoptosis and Caspase-Glo<sup>®</sup> Assay***



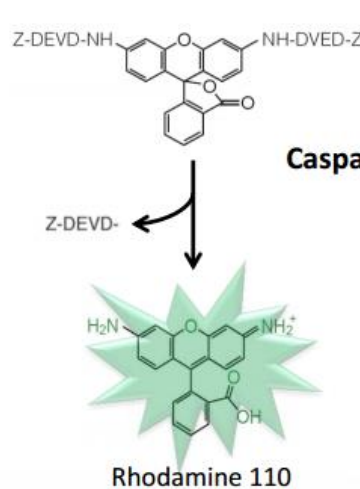
# Apoptosis and Caspase-Glo<sup>®</sup> Assay Range



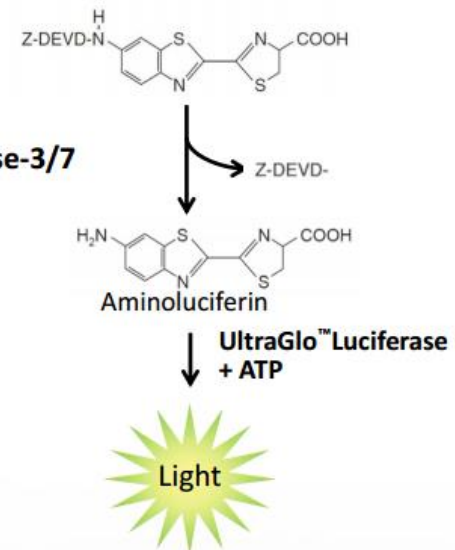
## Promega Assays:

- Caspase-Glo<sup>®</sup> 2
  - **Caspase-Glo<sup>®</sup> 3/7**
  - Caspase-Glo<sup>®</sup> 6
  - Caspase-Glo<sup>®</sup> 8
  - Caspase-Glo<sup>®</sup> 9
- } Luminescence
- **Apo-ONE<sup>®</sup> Caspase 3/7**
- } fluorescence

### Apo-ONE<sup>®</sup> Caspase3/7



### Caspase-Glo<sup>®</sup> Caspase 3/7



## Consensus tetrapeptides:

- VDAD Caspase 2
- **DEVD Caspase 3/7**
- VEID Caspase 6
- LETD Caspase 8
- LEHD Caspase 9

# Apoptosis and Caspase-Glo® Assay Range

## Apo-ONE® Caspase3/7



Thaw and mix the Caspase Substrate and Apo-ONE® Caspase-3/7 Buffer to make the Apo-ONE® Caspase-3/7 Reagent.



Add Apo-ONE® Caspase-3/7 Reagent to each well of a white or black multiwell plate containing blank, control or assay samples.



Gently mix contents of wells using a plate shaker at 300–500rpm for at least 30 seconds. Incubate 30 minutes to 18 hours at room temperature.



Measure fluorescence of each well.

38866A06\_1A

## Caspase-Glo® 3/7 Assay



Add equal volume of reagent to samples.

Mix. Incubate ~30 minutes to 3 hours.



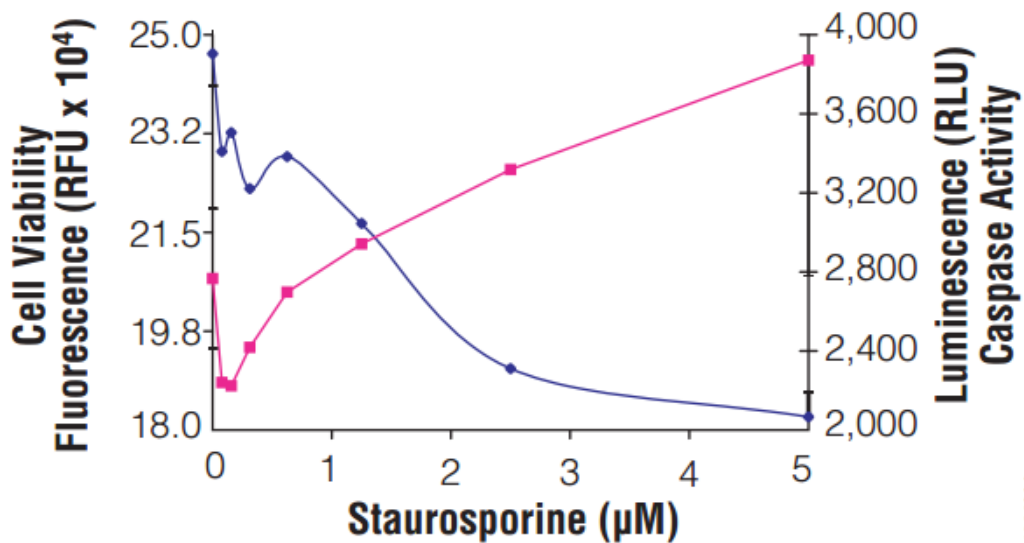
Luminometer

Measure luminescence.

4059A03\_3A

# Apoptosis Detection and Caspase-Glo<sup>®</sup> Assay

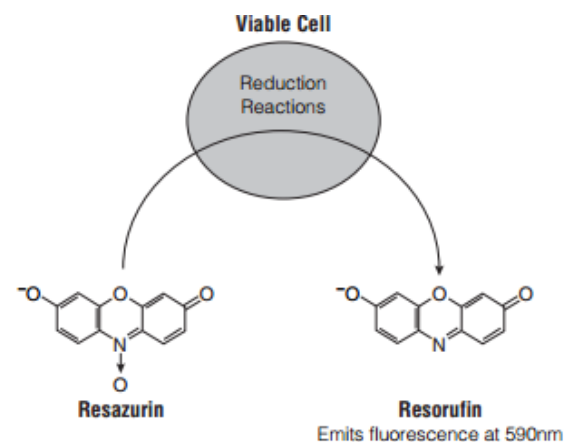
- ◆ CellTiter-Blue<sup>®</sup> Assay signal
- ◆ Caspase-Glo<sup>®</sup> 3/7 Assay signal



HEK293 cells

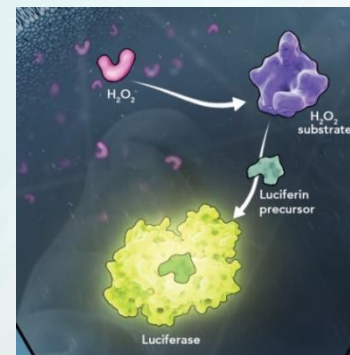
4988MA

## CellTiter-Blue<sup>®</sup> Viability Assay



## ***Cytotoxicity Detection:***

## ***Oxidative Stress and ROS-Glo<sup>®</sup> H<sub>2</sub>O<sub>2</sub> Assay***

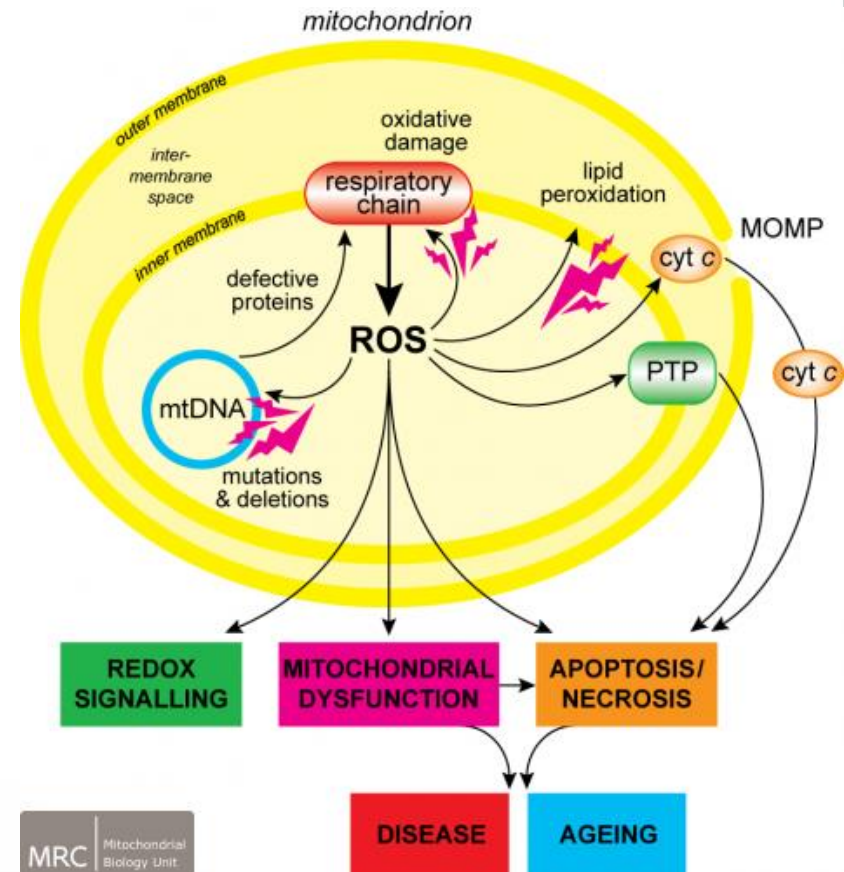


# Cytotoxicity and Oxidative Stress

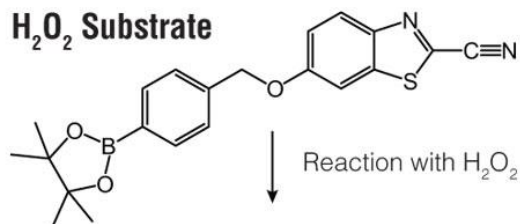
**Oxidative stress:** an imbalance between the production of reactive oxygen species (**ROS**) and the cell's capacity to detoxify the ROS or to repair the oxidative damage.

## Markers of oxidative stress:

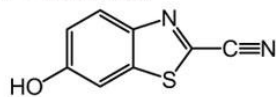
- Altered GSH:GSSG ratio (lowered GSH, increased GSSG)
- **ROS** (super oxide, hydroxyl radical, nitric oxide, hypochlorite convert to more stable  $H_2O_2$ )



# ROS-Glo™ Measures H<sub>2</sub>O<sub>2</sub> Level

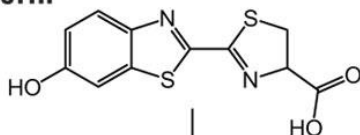


**Luciferin Precursor**



ROS-Glo™ Detection Solution

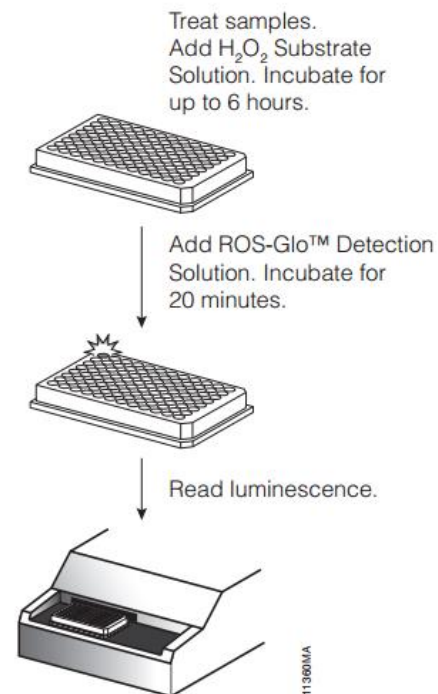
**Luciferin**



11359MB

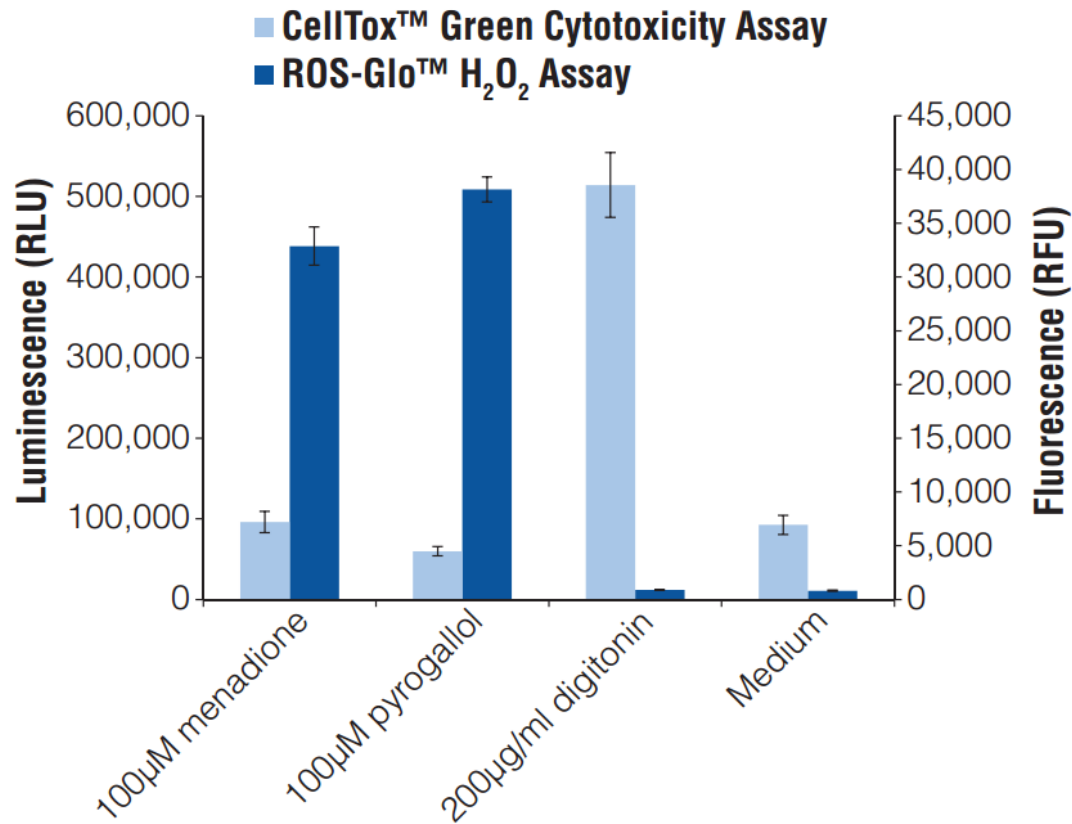
## ROS- Glo™ H<sub>2</sub>O<sub>2</sub> Assay:

- Direct H<sub>2</sub>O<sub>2</sub> detection
- Homogeneous bioluminescent assay ('add-mix-read')



11360MA

# ROS-Glo™ H<sub>2</sub>O<sub>2</sub> Assay

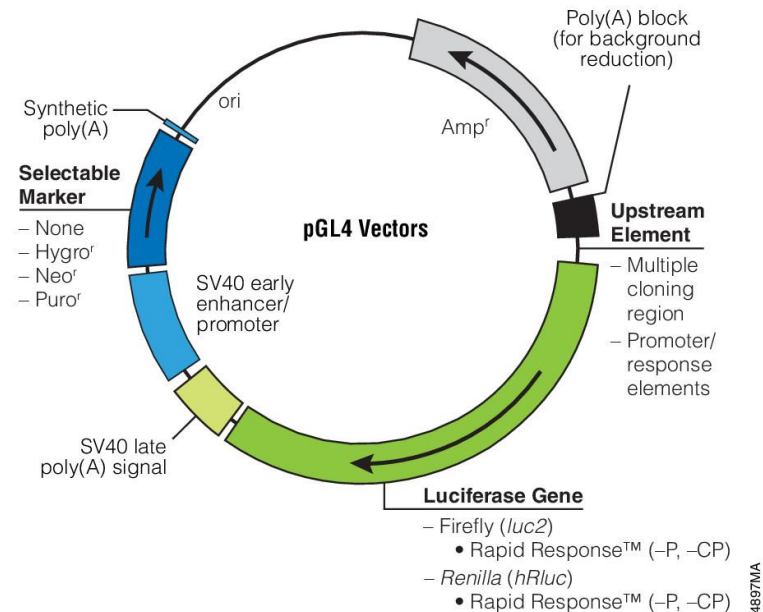


11586MA

HepG2 cells

# pGL4 Firefly Vectors for Signalling Pathway Analysis

- Vectors used to create a cell line with an indicator of a functional signaling pathway
- Useful for deciphering the components in a signaling pathway
- Made by insertion of multiple repeats of a response element upstream of a minimal promoter (minP)
- Collection of vectors that report the activity of a variety of pathways using the optimized **luc2 firefly luciferase gene** in the **pGL4 backbone**
- **Hygromycin resistance** selectable marker, allowing use either in transient transfection experiments or for selection of a stable cell line



# *pGL4 Firefly Vectors for Signaling Pathway Analysis*

Activator/Pathway	Transcription Factor	Binding Site	Vector Name	Catalogue
Oxidative Stress	Nrf2	Antioxidant Response Element (ARE)	pGL4.37[ <i>luc2P</i> /ARE/Hygro]	E3641
DNA Damage	p53	p53 Response Element (p53 RE)	pGL4.38[ <i>luc2P</i> /p53 RE/Hygro]	E3651
Heavy Metal Stress	MTF1	Metal Regulatory Element (MRE)	pGL4.40[ <i>luc2P</i> /MRE/Hygro]	E4131
Heat Shock	HSF1	Heat Shock Element (HSE)	pGL4.41[ <i>luc2P</i> /HSE/Hygro]	E3751
Hypoxia	Hif1 $\alpha$	Hypoxia Response Element (HRE)	pGL4.42[ <i>luc2P</i> /HRE/Hygro]	E4001
Xenobiotic Stress	AhR	Xenobiotic Responsive Element (XRE)	pGL4.43[ <i>luc2P</i> /XRE/Hygro]	E4121
Endoplasmic Reticulum Stress/ Unfolded Protein Response	ATF6	Activating Transcription Factor 6 Response Element (ATF6 RE)	pGL4.39[ <i>luc2P</i> /ATF6 RE/Hygro]	E3661

# ***Cytotoxicity Assays***

## **Cell Viability Assays:**

- RealTime-Glo™ MT
- CellTiter-Glo® 2.0
- CellTiter-Glo® 3D
- CellTiter-Fluor™
- CellTiter-Blue™
- CellTiter96®-AQueous
- BacTiter-Glo® Microbial

## **Apoptosis Assays:**

- Caspase-Glo® 2
- Caspase-Glo® 3/7
- Caspase-Glo® 6
- Caspase-Glo® 8
- Caspase-Glo® 9
- Apo-ONE® Caspase 3/7
- CaspACE™ Assay System
- DeadEnd™ Fluorometric TUNEL
- DeadEnd™ Colorimetric TUNEL

## **Cytotoxicity Assays:**

- CellToxGreen™
- CytoTox-Glo™
- CytoTox-Fluor™
- CytoTox-ONE™
- CytoTox™ 96

## **Metabolism Assays:**

- NAD/NADH-Glo™
- NADP/NADPH-Glo™
- NAD(P)H-Glo™ Detection System

## **Oxidative Stress Assays:**

- ROS-Glo™ H<sub>2</sub>O<sub>2</sub> Assay
- GSH-Glo™ Glutathione Assay
- GSH-GSSG-Glo™ Assay

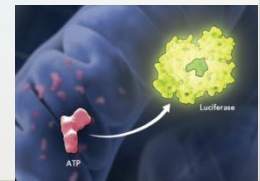
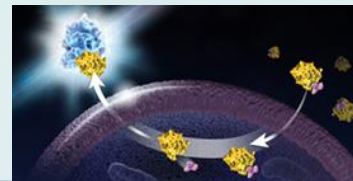
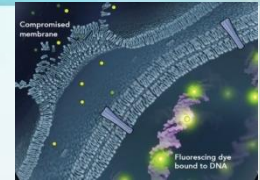
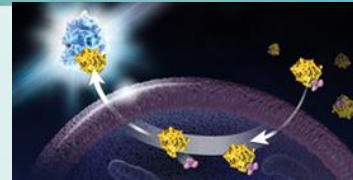
## **Mitochondrial Toxicity Assays:**

- Mitochondrial ToxGlo™

## **Viral Toxicity Assays:**

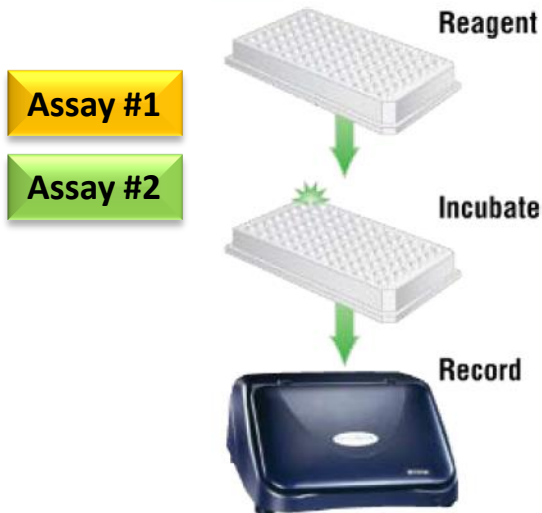
- Viral-ToxGlo™ Assay

# *Multiplexing*



# Multiplexing Set-up

continuous assay #1  
and  
continuous assay #2



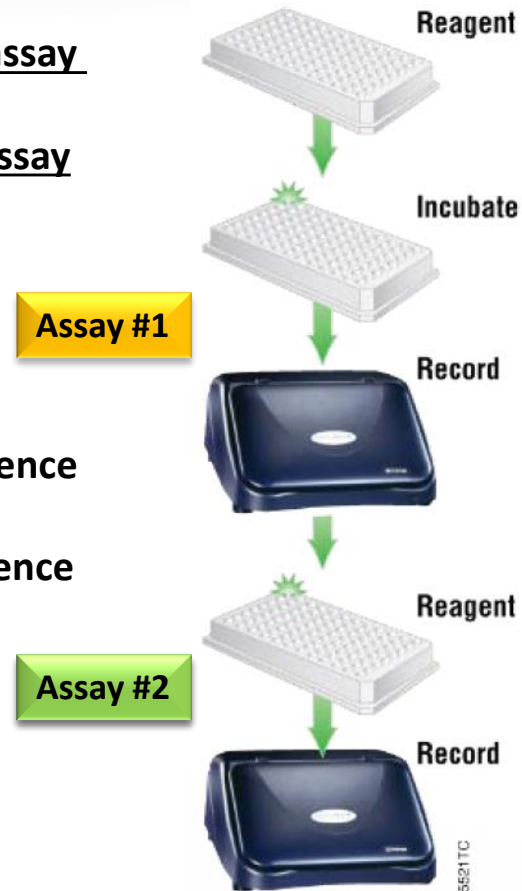
RealTime-Glo™ MT Cell Viability Assay

CellTox™ Green Cytotoxicity Assay

luminescence + fluorescence

continuous assay  
and  
end-point assay

luminescence  
+  
fluorescence

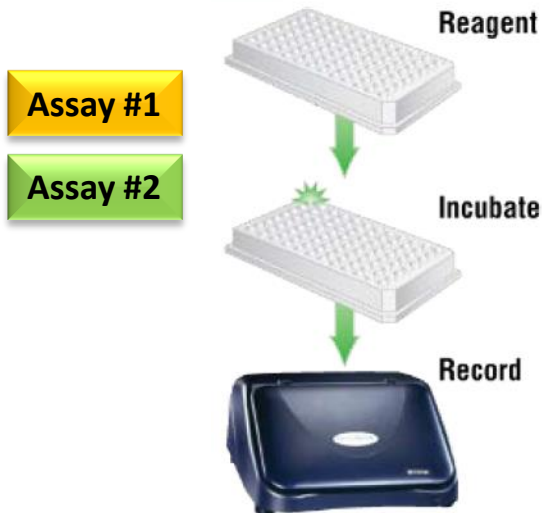


RealTime-Glo™ MT Cell Viability Assay

CytoTox-Fluor™ Viability Assay

# Multiplexing Set-up

continuous assay #1  
and  
continuous assay #2

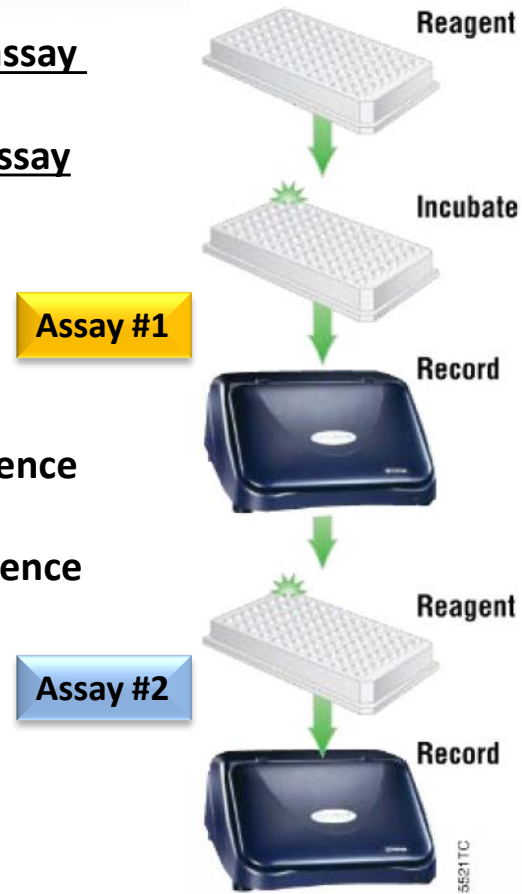


RealTime-Glo™ MT Cell Viability Assay

CellTox™ Green Cytotoxicity Assay

luminescence + fluorescence

continuous assay  
and  
end-point assay



luminescence  
+  
luminescence

RealTime-Glo™ MT Cell Viability Assay

CellTiter-Glo® Cell Viability Assay

# Multiplexing - Examples

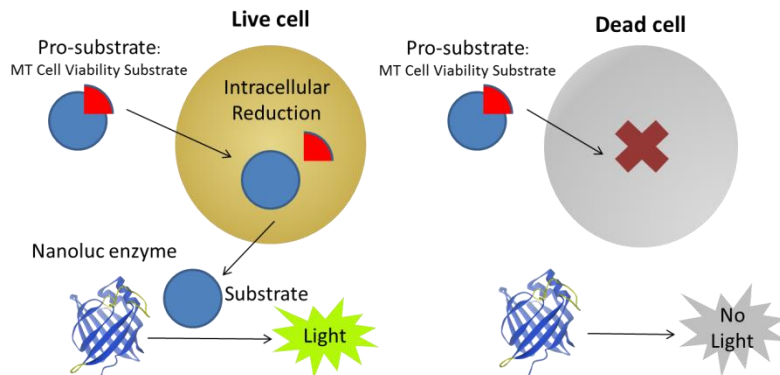
**Table 1. Assays that can be Multiplexed with the RealTime-Glo™ MT Cell Viability Assay.**

<b>First Assay</b>	<b>Second Assay</b>	<b>Information Obtained by Multiplexing</b>
RealTime-Glo™ MT Cell Viability Assay	CellTox™ Green Cytotoxicity Assay <sup>1</sup>	Cell viability and cytotoxicity (membrane integrity)
RealTime-Glo™ MT Cell Viability Assay	CytoTox-Fluor™ Cytotoxicity Assay	Cell viability and cytotoxicity (protease release)
RealTime-Glo™ MT Cell Viability Assay	CellTiter-Glo® Luminescent Cell Viability Assay	Two independent markers of cell viability (reducing potential and ATP)
RealTime-Glo™ MT Cell Viability Assay	NAD/NADH-Glo™ Assay	Cell viability and measurement of NAD <sup>+</sup> and NADH
RealTime-Glo™ MT Cell Viability Assay	NADP/NADPH-Glo™ Assay	Cell viability and measurement of NADP <sup>+</sup> and NADPH
RealTime-Glo™ MT Cell Viability Assay	Reporter assays <sup>2</sup>	Cell viability and reporter gene activity
RealTime-Glo™ MT Cell Viability Assay	RNA isolation	Cell viability and RNA analysis

# Multiplexing and Mechanism of Cytotoxicity (1)

## RealTime-Glo™ MT Cell Viability Assay

### continuous assay #1

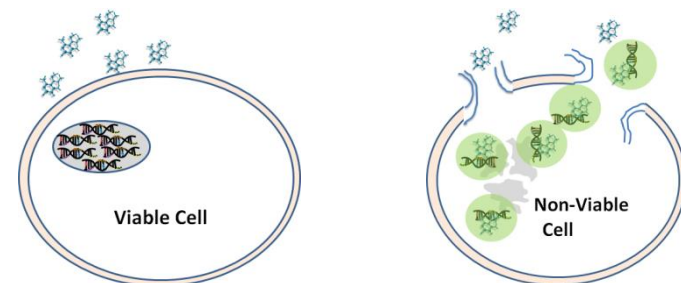


- **Luminescent**
- Metabolically active cells
- **Viable cells → high signal**
- Dead cells → low signal



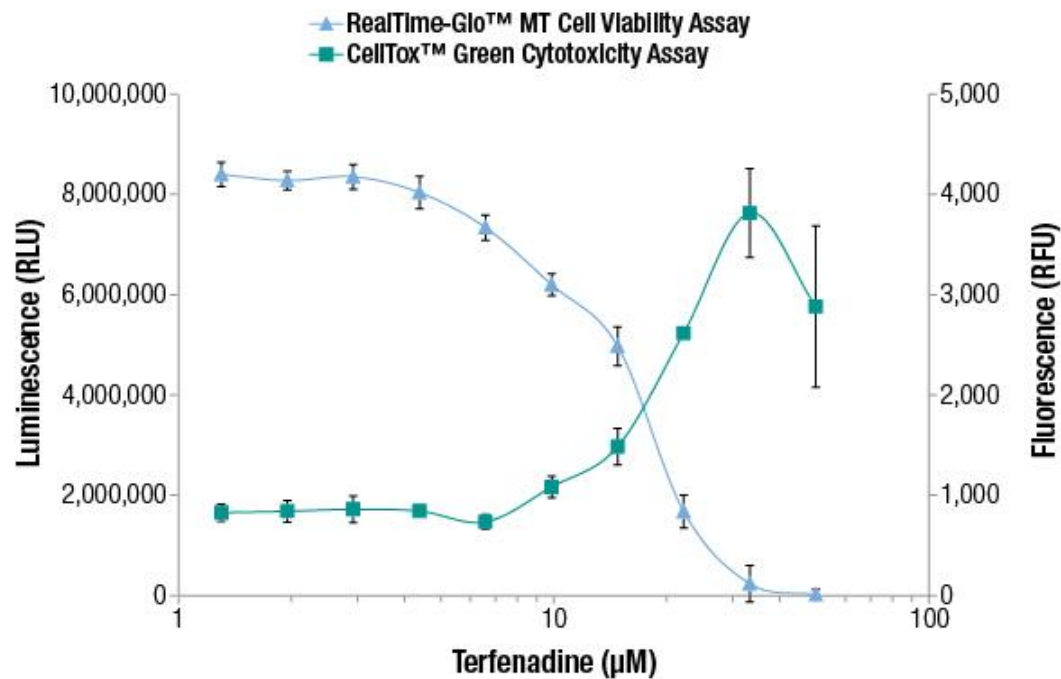
## CellTox™ Green Cytotoxicity Assay

### continuous assay #2



- **Fluorescent**
- DNA released from dead cells
- **Viable cells → low signal**
- Non-viable cells → high signal

# Multiplexing and Mechanism of Cytotoxicity (1)

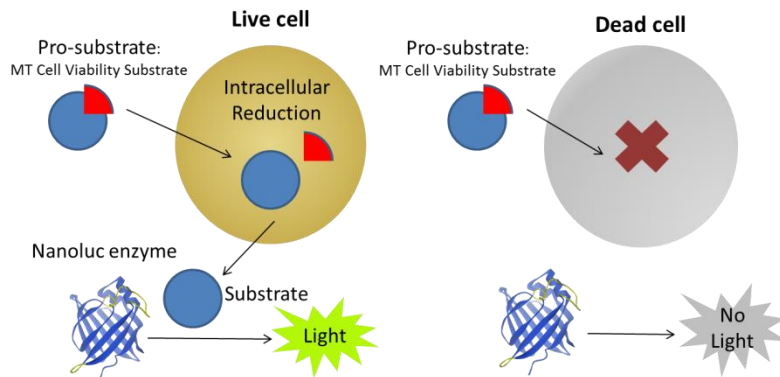


Hep2G frozen instant cells; 10,000 cells/ well  
 96-well white plate; 6h time- point

# Multiplexing and Mechanism of Cytotoxicity (2)

## RealTime-Glo™ MT Cell Viability Assay

### continuous assay #1

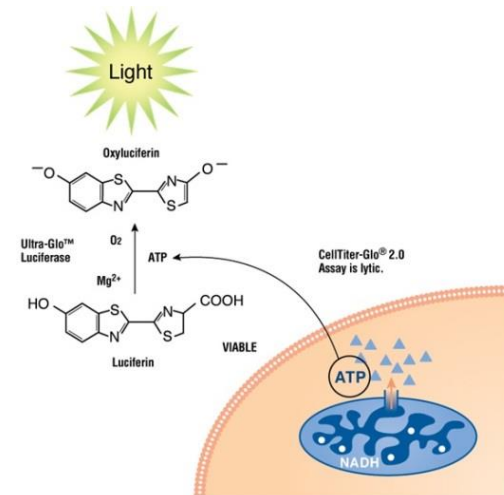


- **Luminescent**
- Metabolically active cells
- **Viable cells → high signal**
- Dead cells → low signal



## CellTiter-Glo® Cell Viability Assay

### end point assay #1

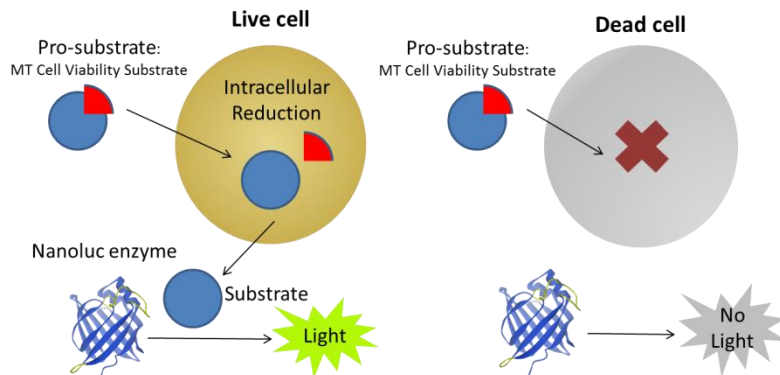


- **Luminescent**
- ATP
- **Viable cells → high signal**
- Dead cells → low signal

# Multiplexing and Mechanism of Cytotoxicity (3)

## RealTime-Glo™ MT Cell Viability Assay

continuous assay #1



- **Luminescent**
- Metabolically active cells
- **Viable cells → high signal**
- **Dead cells → low signal**



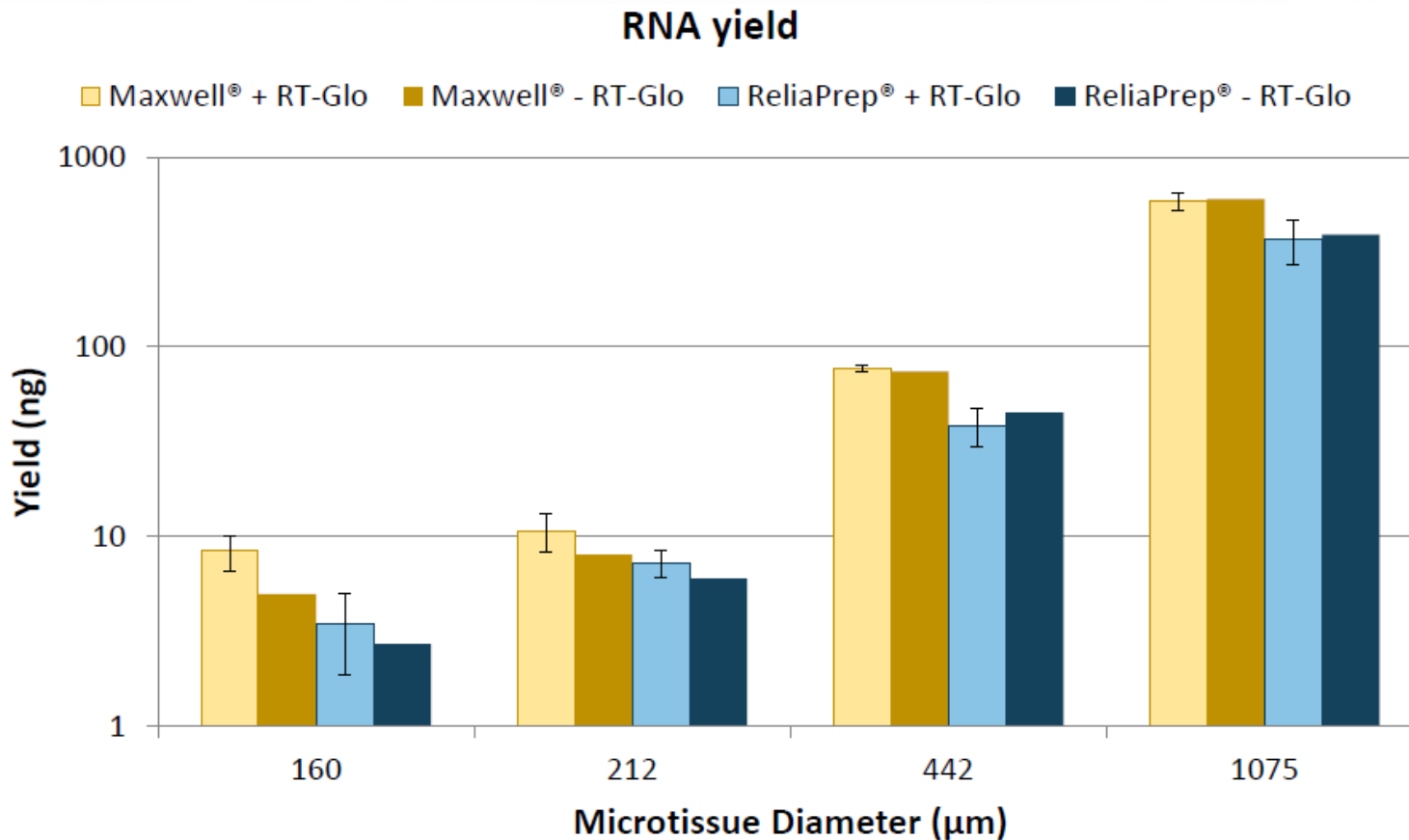
## RNA Isolation

end point assay #1



- Extract RNA using **ReliaPrep™ Assay**
- Perform qPCR
- Gene expression microarray
- RNA Seq

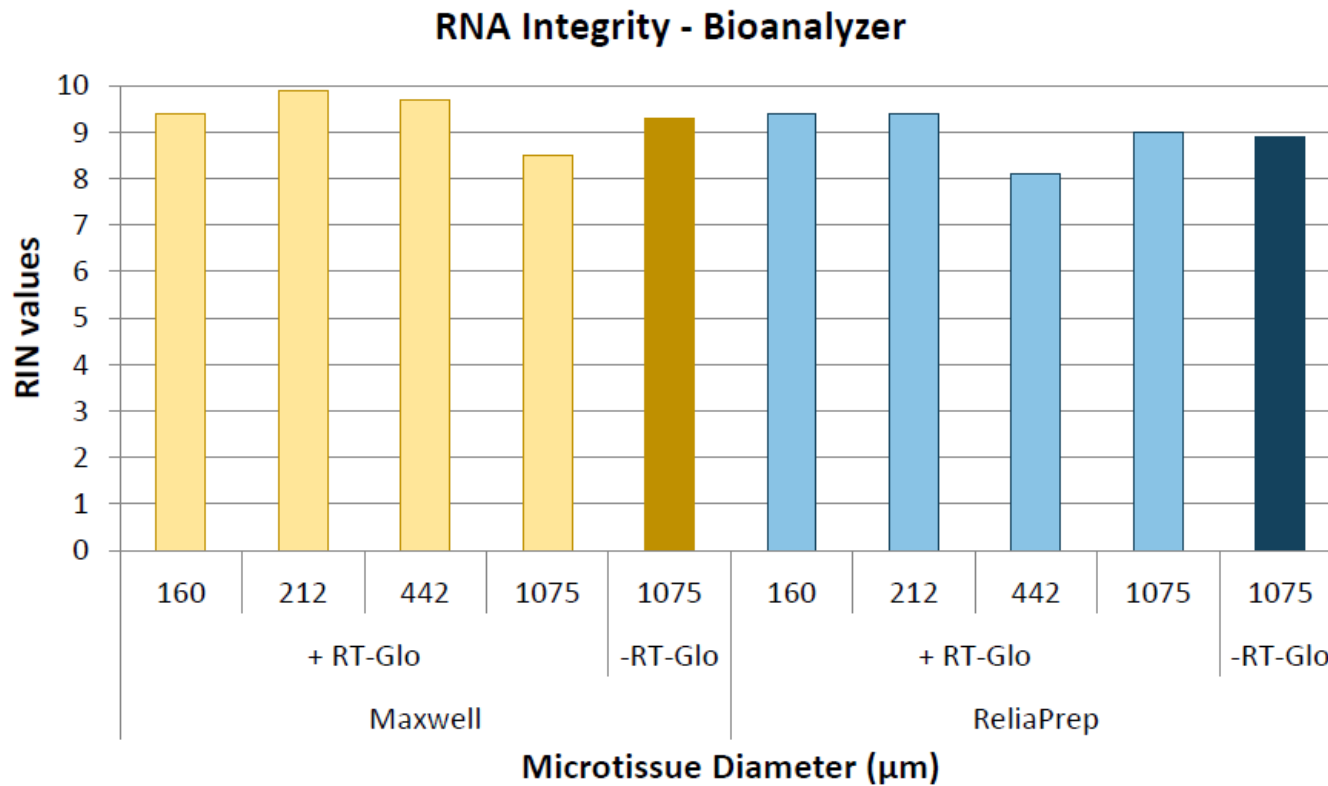
# Multiplexing and Mechanism of Cytotoxicity (3)



**RealTime-Glo™ reagents do NOT effect**  
**RNA yield using QuantiFluor® RNA System**

# Multiplexing and Mechanism of Cytotoxicity (3)

RNA integrity is NOT affected by presence of RealTime-Glo™ reagents!



## ***Multiplexing and Mechanism of Cytotoxicity (3)***

**RealTime-Glo™ Reagent does NOT affect cycle threshold of extracted RNA**

**Table 2.  $C_t$  Values Generated from RNA Isolated from Different Cell Types With or Without the RealTime-Glo™ Reagent.**

Cell Type	$C_t$ Value	
	With Medium Only	With RealTime-Glo™ Reagent
A549	30.9	30.3
K562	31.3	30.2
MCF7	30.9	30.1
THP-1	29.9	29.9

# Summary

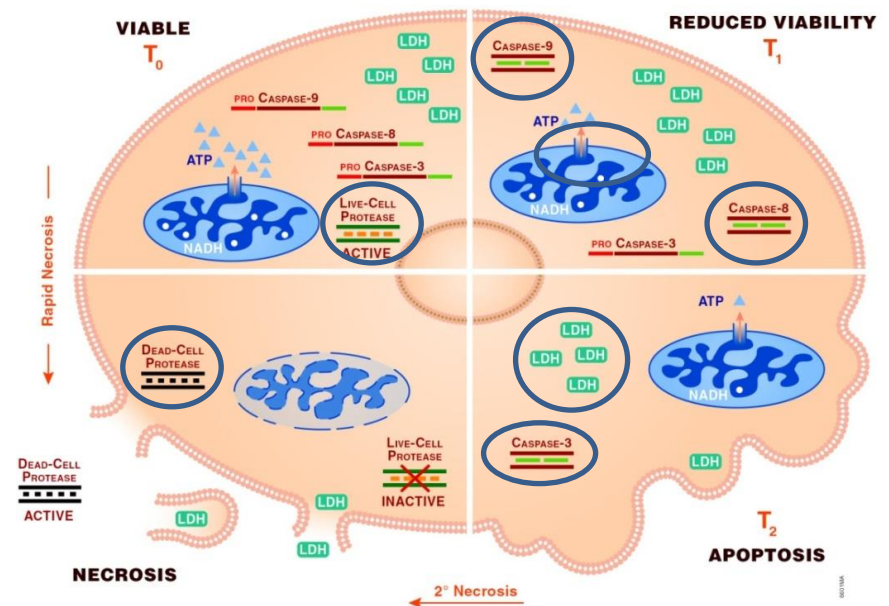
## Multiplexing gives a more complete picture of what's happening in the cell

### Requirements :

- Assays must be biologically & chemically compatible
- Signals must be spectrally different

### Advantages:

- Reduces ambiguity
- Eliminates variables
- Normalises data
- Increases data content/well/ aliquot



# Conclusions

## Real-Time Glo™ /CellToxGreen™:

- Provides kinetic information about viable/ dead cell number during the course of the experiment that enables 'on the go' decision making
- Reagents are non- toxic to the cells
- Optional protocols enables adding the reagents at different stages of the experiment
- **Multiplexing** opportunities with other assays to determine **the mechanism of cytotoxicity** and possible involvement of other processes such as oxidative stress