

Improved T Cell Activation Bioassays for Development of Bispecific Antibodies and Engineered T Cell Immunotherapies

Pete Stecha, Denise Garvin, Jim Hartnett, Frank Fan, Mei Cong and Zhi-jie Jey Cheng

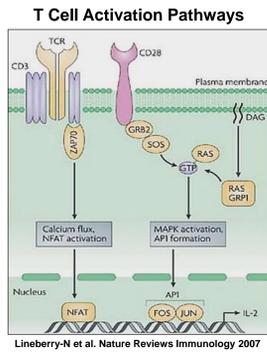
Promega Corporation, 2800 Woods Hollow Rd, Madison, WI 53711



1. Introduction

Immunotherapy aims to boost a patient's own immune system to fight disease. The strategy is aimed at inducing, strengthening or engineering T cell responses for the treatment of cancer and autoimmune disease.

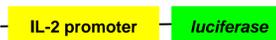
Here we describe a platform of T cell activation bioassays for the development of CD3 bispecific antibodies and engineered T cell immunotherapies. Specifically, we developed two bioluminescent reporter-based bioassays to measure T cell activation via TCR/CD3 or TCR/CD3 plus CD28 co-stimulation.



TCR/CD3 (NFAT) effector cells: Jurkat cells engineered with an NFAT-RE driving luciferase expression. Responds to TCR/CD3, but not CD28 stimulation.

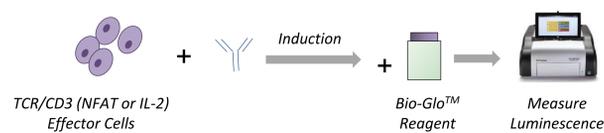


TCR/CD3 (IL-2) effector cells: Jurkat cells engineered with an IL-2 promoter driving luciferase expression. Responds to TCR/CD3 and CD28 stimulation.

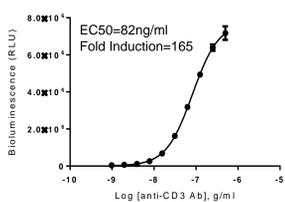


2. TCR/CD3 (NFAT or IL-2) Effector Cells Respond to TCR/CD3 Stimulation

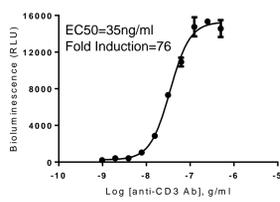
T Cell Activation Protocol



TCR/CD3 (NFAT)

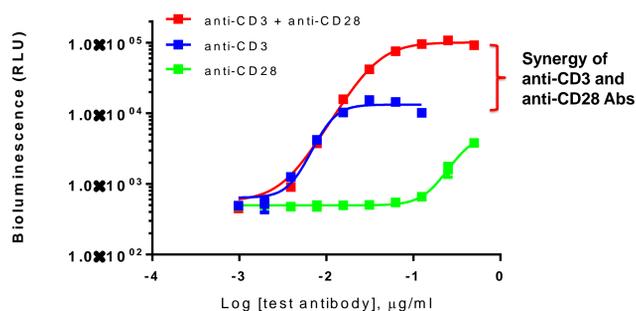


TCR/CD3 (IL-2)



TCR/CD3 (NFAT) (Left) or TCR/CD3 (IL-2) (Right) effector cells were stimulated with increasing concentrations of an anti-CD3 antibody and secondary crosslinking antibody.

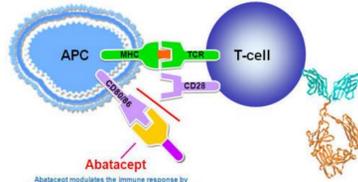
3. TCR/CD3 and/or CD28 Co-stimulation Measured using TCR/CD3 (IL-2) Effector Cells



Legend for TCR/CD3 (IL-2) effector cells:
 - anti-CD3 + anti-CD28 Abs: EC50=36ng/ml, Fold Induction=230
 - anti-CD3 Ab: EC50=11ng/ml, Fold Induction=31
 - anti-CD28 Ab: EC50=>368ng/ml, Fold Induction=8

TCR/CD3 (IL-2) effector cells were stimulated with increasing concentrations of anti-CD28, anti-CD3 or a combination of anti-CD3+anti-CD28 Abs, as indicated.

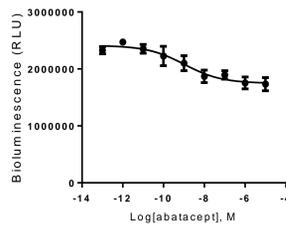
4. Abatacept Inhibits CD28 Co-stimulation in TCR/CD3 (IL-2) Effector Cells



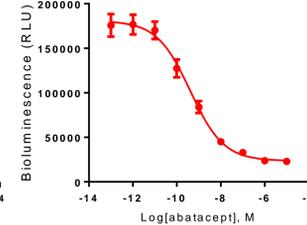
Experimental Design

- (1) TCR/CD3 (NFAT or IL-2) effector cells are incubated with Raji (CD80/86⁺) target cells.
- (2) T cell activation is induced via crosslinked anti-CD3 Ab and CD28 engagement by its ligand CD80/86 expressed on the Raji target cells.
- (3) Addition of a CTLA-4/IgG fusion protein (Abatacept) binds CD80/86 and inhibits CD28-mediated T cell activation.

TCR/CD3 (NFAT)



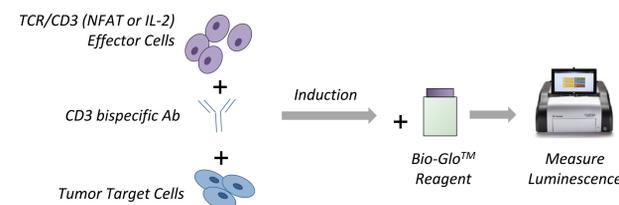
TCR/CD3 (IL-2)



Increasing concentrations of Abatacept were added to either TCR/CD3 (NFAT) or TCR/CD3 (IL-2) effector cells, as indicated. Abatacept induced a significant decrease in TCR-mediated luciferase activity in TCR/CD3 (IL-2) effector cells compared to TCR/CD3 (NFAT) effector cells. This is expected because CD28 functions independently of the NFAT response element (see Introduction).

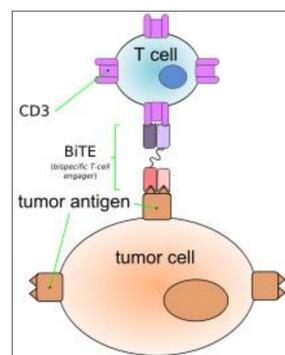
5. Analysis of CD3xCD19 Bispecific Antibody Blinatumomab Activity

Assay Protocol for Measuring CD3 Bispecific Antibody Activity



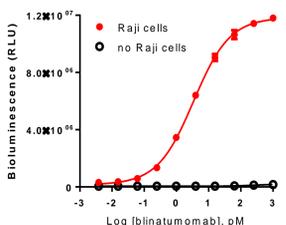
Experimental Design

- (1) TCR/CD3 (NFAT or IL-2) effector cells are incubated with increasing concentrations of a CD3 bispecific Ab
- (2) The bispecific Ab simultaneously binds to TCR/CD3 on the effector cells and tumor antigen on the target cells
- (3) Bispecific Ab binding stimulates IL-2 or NFAT luciferase activity

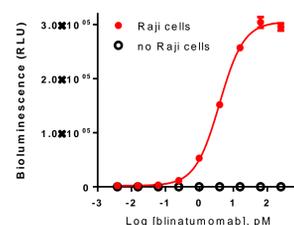


Blinatumomab induced a dose-dependent increase in luciferase activities in both TCR/CD3 (IL-2) and TCR/CD3 (NFAT) effector cells in the presence of Raji (CD19⁺) target cells. No response was detected in the absence of Raji (CD19⁺) target cells.

TCR/CD3 (NFAT)



TCR/CD3 (IL-2)



6. Assay Qualification with Blinatumomab: Assay Precision, Accuracy, and Linearity

Accuracy and Intermediate Precision (N = 6)

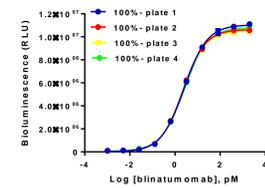
Assay Qualification Design:

- Two analysts
- Three days
- Four plates per day
- 100% vs 50%
- 100% vs 75%
- 100% vs 150%
- 100% vs 200%

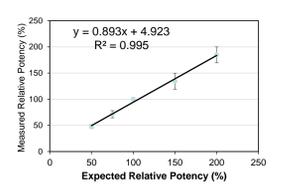
Data shown are generated using TCR/CD3 (NFAT) effector cells, blinatumomab, and Raji (CD19⁺) target cells.

Expected Relative Potency	Assay	Analyst	Measured Relative Potency %	Mean %	SD%	Accuracy %	Precision RSD %
50%	1	1	45.3	47.8	3.6	95.7	7.6
	2	1	45.0				
	3	1	45.5				
	4	2	46.2				
	5	2	52.2				
	6	2	52.8				
75%	1	1	62.2	71.2	6.8	95.0	9.5
	2	1	73.9				
	3	1	63.3				
	4	2	78.4				
	5	2	75.9				
	6	2	73.6				
150%	1	1	144.5	134.4	15.5	89.6	11.5
	2	1	143.5				
	3	1	121.4				
	4	2	108.8				
	5	2	143.4				
	6	2	144.7				
200%	1	1	174.9	184.8	15.0	92.4	8.1
	2	1	195.5				
	3	1	179.8				
	4	2	162.0				
	5	2	198.9				
	6	2	198.0				
Overall				93.2		9.2	

Repeatability (%CV) = 3.01%

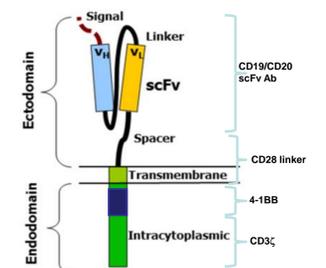


Linearity and Range (R²=0.995)

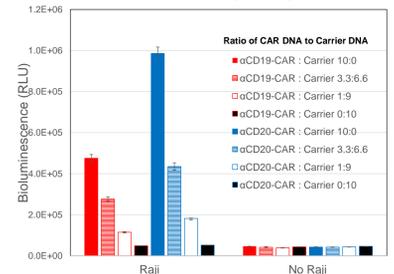


7. Measurement of CAR-T Cell Activity

CAR-T Design



TCR/CD3 (NFAT)



TCR/CD3 (NFAT) effector cells were transiently transfected with increasing amounts of DNA for anti-CD19- or anti-CD20-chimeric antigen receptors (CD19/CARs or CD20/CARs). The resulted CAR-T effector cells were incubated in the presence or absence of Raji (CD19/CD20⁺) target cells. Luciferase activity was detected from CAR-T effector cells in the presence of Raji cells, but not from CAR-T effector cells alone.

Similar data were generated using TCR/CD3 (IL-2) effector cells (data not shown)

8. Conclusions

We have developed a platform of T cell activation bioassays that incorporate a bioluminescent reporter-based readout of T cell activation via NFAT or IL-2 promoter.

These assays reflect the Mechanism of Action (MoA) of biologics designed to engage and stimulate T cell activation to attack target disease cells, and provide consistent and reliable measurement of potency of pathway activation for anti-CD3 bispecific Ab and CAR-T cell activity.

Easy-to-implement with a rapid and convenient workflow that is amenable to standard 96-well and 384-well plate formats. All assays can be used in "thaw-and-use" cell format, no cell culture required and are suitable for development into potency, stability, and NAb assays.