

What transfection reagent should I choose?

Promega offers three premium transfection reagents: ViaFect[™], FuGENE[®] 6 and FuGENE[®] HD. All three offer broad spectrum transfection capability and low toxicity but the same question always comes up: Which one should I choose? First, not every transfection reagent is perfect for every cell line. Second, transfection success can also vary with DNA quality, passage number of cells, cell density, etc.

To get a better understanding of how each reagent performs on a variety of cell lines, a transfection trial was performed with the three Promega reagents and the leading competitor's transfection reagent . Each reagent was tested with a range of transfection reagent: DNA ratios and two concentrations of DNA.

Experiments

See page 2. Cells were transfected with either 50ng or 100ng of luciferase reporter plasmid at various reagent:DNA ratios.

Results

A summary of the results are presented in Table 1 with more information on the exact best conditions in Table 2.

The leading competitor and the Promega reagents suggest trying a 3:1 ratio with 100ng of DNA per well of a 96-well plate if you need a starting place. Table 3 demonstrates how the reagents perform relative to one another when you use the common conditions.

Conclusion

All four transfection reagents work on a variety of cells. Each reagent clearly works best on certain cell lines but none were perfect for every cell line. Table 2 demonstrates that the common 3:1 ratio is not always the most optimal condition. ViaFect™ Reagent yielded greater luciferase expression over a wider variety of cells in this test whether you optimize or just choose one condition.

This summary is based on data generated by Brad Hook and Alisha Truman, Promega Scientific Applications Services, June 2014.

Table 1. Summary of Transfection Study.Transfection was assessed through expression of fireflyluciferase (EXP). The best condition (highest RLUs) was chosen for each transfection reagent. For each cell line, the maximum RLUs were set at 100% for one reagent and the other 3 were judged relative to that reading. Cells were also assayed for cell viability (**VIA**) and judged versus a no-transfection control for each cell line. Transfection efficiencies lower than 50% are not reported on this table. Greater detail is reported in Table 2.

Cell Line		FuGENE [®] 6		FuGENE [®] HD		ViaFect™ Reagent		Competitor L2K	
		EXP	VIA	EXP	VIA	EXP	VIA	EXP	VIA
A549)	++	++	+++	+++	+++	+++		
C2C12						+++	+++		
СНО ++ +++		+++			+++	++	++	++	
cos	7	+++	+++	+++	+++	+++	+++	+++	+++
H9C	2					+++	+++	++	+++
HCT116				+++	++	+++	+++	++	++
HEK 293		++	++			+++	+++		
HeLa				+++	+++	++	+++		
HepG2						+++	++		
HT-29						+++	+++	+++	+++
Huh	7					+++	++	++	++
Jurk	at					+++	+++	++	+++
K562	2							+++	+++
LNC	aP					+++	+++		
MCF	7					+++	+++	+++	+++
NIH	3T3					++	+++	+++	+++
PC-12				++	+++	+++	+++	+++	+++
PC-3				+++	+++	+++	+++		
RAW 264.7		+++	++	+++	++			++	++
U2OS				++	+++	+++	+++		
KEY	+++	• >80% • >80% viable	of maxR of cells v	LUs; vere	++	• >50% to <80% max RLUs; • >50% to <80% of cells were viable			



Experimental Overview

 Transfection reagents were prepared at room temperature following the directions for each transfection reagents. The following reagent to DNA ratios were tested:

FuGENE [®] 6 Reagent	6:1	4:1	3:1	1.5:1
FuGENE [®] HD Reagent	4:1	3:1	2.5:1	2:1
ViaFect™ Reagent	4:1	3:1	2.5:1	2:1
Competitor L2K	5:1	4:1	3:1	2:1

- Each reagent:DNA ratio was pipetted onto 6 wells of cells in either 5µl or 10µl volumes. One 96 well plate/transfection reagent was used per cell line.
- 3. Plates were mixed for 15 seconds on a plate shaker for 10-15 seconds
- Plates were transferred to a 37°C/5% CO₂ incubator for 24 hours prior to assay for cell viability and luciferase reporter activity. Wells with cells only were 100% viability controls.

More complete experimental details are available upon request.







Table 2. Optimal 96-well transfection conditions for each cell line at 24 hours post-transfection.

24 nours post-transfection. Transfection was judged through expression of firefly luciferase. The conditions that produced the highest RLUs for each transfection reagent were compared relative to the reagent that produced the highest overall RLUs (MAX). The percent of RLU Max is reported along with the optimal reagent:DNA ratio and whether 50ng (5µl) or 100ng (10µl) produced the highest RLUs for that reagent. Viability was judged relative to untransfected control cells. Details for luciferase expression below 50% of MAX RLUs are not reported in this table.

Cell Line	Measure	FuGENE [®] 6	FuGENE [®] HD	ViaFect™	L2K	
AE 40	% RLU Max (ratio; vol.)	60% (4:1; 10µl)	90% (3:1; 5µl)	MAX (4:1; 10µl)	30%	
A549	% Viable @ 24hr	>90%	>90%	>90%	60%	
00040	% RLU Max (ratio; vol.)	<10%	<10%	MAX (4:1; 10µl)	25%	
62612	% Viable @ 24hr	>90%	>90%	>90%	>90%	
cuo	% RLU Max (ratio; vol.)	60% (6:1; 5µl)	30%	MAX (4:1; 10µl)	65% (2:1; 10µl)	
СПО	% Viable @ 24hr	>90%	>90%	65%	60%	
0007	% RLU Max (ratio; vol.)	MAX (3:1; 10µl)	90%(3:1; 5µl)	80% (4:1; 10µl)	90% (4:1; 10µl)	
0037	% Viable @ 24hr	>90%	>90%	>90%	85%	
H0C2	% RLU Max (ratio; vol.)	10%	<10%	MAX (4:1; 10µl)	50% (3:1; 10µl)	
HIGCZ	% Viable @ 24hr	>90%	>90%	>90%	85%	
	% RLU Max (ratio; vol.)	15%	MAX (2.5:1; 5µl)	95% (2:1; 5µl)	60% (2:1; 5µl)	
HC1116	% Viable @ 24hr	80->90%	70%	80%	70%	
	% RLU Max (ratio; vol.)	60% (4:1; 10µl)	20%	MAX (4:1; 10µl)	30%	
HER 295	% Viable @ 24hr	75%	60%	>90%	75%	
	% RLU Max (ratio; vol.)	30%	MAX (3:1; 5µl)	60% (4:1; 10µl)	45%	
песа	% Viable @ 24hr	80%	80%	>85%	70%	
HonG2	% RLU Max (ratio; vol.)	30%	30%	MAX (4:1; 10µl)	30%	
перөг	% Viable @ 24hr	75%	55%	55%	65%	
	% RLU Max (ratio; vol.)	<10%	40%	MAX (3:1; 10µl)	90% (3:1; 10µl)	
11-29	% Viable @ 24hr	>90%	>90%	>90%	>90%	
Hub7	% RLU Max (ratio; vol.)	40%	10%	MAX (2.5:1; 5µl)	60% (2:1; 5µl)	
Hull7	% Viable @ 24hr	80%	75%	75%	70%	
Jurkat	% RLU Max (ratio; vol.)	<10%	<10%	MAX (2:1; 5µl)	65% (2:1; 10µl)	
	% Viable @ 24hr	>90%	>90%	>90%	80%	
K562	% RLU Max (ratio; vol.)	<10%	10%	30%	MAX (2:1; 10µl)	
	% Viable @ 24hr	>90%	>90%	>90%	>90%	
	% RLU Max (ratio; vol.)	<10%	40%	MAX (4:1; 10µl)	20%	
	% Viable @ 24hr	>90%	75%	>90%	50%	
MCE7	% RLU Max (ratio; vol.)	10%	15%	80% (4:1; 10µl)	MAX (2:1; 10µl)	
	% Viable @ 24hr	>90%	>90%	>90%	80%	
NIH 3T3	% RLU Max (ratio; vol.)	<10%	15%	60% (4:1; 10µl)	MAX (4:1; 10µl)	
	% Viable @ 24hr	>90%	>90%	>90%	>90%	
PC-12	% RLU Max (ratio; vol.)	10%	70% (2.5: 1; 10µl)	MAX (4:1; 10µl)	90% (2:1; 10µl)	
	% Viable @ 24hr	>90%	>90%	>90%	>90%	
PC-3	% RLU Max (ratio; vol.)	15%	90% (4:1; 10µl)	MAX (4:1; 10µl)	15%	
	% Viable @ 24hr	>90%	>90%	80%	40-70%	
RAW 264.7	% RLU Max (ratio; vol.)	MAX (3:1; 10µl)	85% (2.5:1; 5µl)	<10%	50% (2:1; 5µl)	
	% Viable @ 24hr	75%	65%	>90%	70%	
U2OS	% RLU Max (ratio; vol.)	40%	60% (3:1; 5µl)	MAX (4:1; 10µl)	20%	
	% Viable @ 24hr	>90%	>90%	>90%	>90%	
	≥80% of RLU Max Viability ≥80% @ 24hr		50 to <80% of RLU Max >50% to <80% Viability @ 24hr			



Table 3: Comparison of all four transfection reagents at the commonly recommended conditions of 3:1 Reagent:DNA ratio and 100ng DNA per well of a 96-well plate.

This data was extracted from the overall transfection study. Luciferase expression (EXP) is judged for each cell line relative to the maximum RLUs observed (MAX). Viability (VIA; % of untransfected cells) is listed for each cell line under these conditions.

Cell Line	FuGENE [®] 6		FuGl H	FuGENE [®] ViaFect HD Reage		ect™ gent	Competitor L2K	
	EXP	VIA	EXP	VIA	EXP	VIA	EXP	VIA
A549	60%	>90%	60%	60%	MAX	>90%	<10%	30%
C2C12	15%	>90%	<10%	>90%	MAX	>90%	15%	>90%
СНО	55%	>90%	40%	>90%	MAX	85%	50%	45%
COS7	MAX	85%	90%	>90%	60%	>90%	85%	>90%
H9C2	15%	>90%	<10%	>90%	MAX	>90%	65%	>90%
HCT116	20%	80%	80%	65%	MAX	80%	40%	45%
HEK 293	70%	75%	20%	50%	MAX	>90%	30%	40%
HeLa	25%	>90%	МАХ	65%	65	>90%	40%	50%
HepG2	35%	85%	20%	50%	MAX	55%	25%	55%
HT-29	<10%	>90%	45%	>90%	MAX	>90%	90%	85%
Huh7	40%	70%	<10%	40%	MAX	70%	25%	45%
Jurkat	35%	>90%	<10%	>90%	MAX	80%	95%	80%
K562	<10%	>90%	<10%	>90%	20%	>90%	MAX	>90%
LNCaP	<10%	>90%	60%	75%	MAX	>90%	25%	60%
MCF7	<10%	>90%	20%	>90%	95%	>90%	MAX	70%
NIH 3T3	<10%	>90%	25%	>90%	65%	>90%	MAX	>90%
PC-12	15%	>90%	70%	>90%	MAX	>90%	95%	85%
PC-3	20%	>90%	55%	80%	MAX	>90%	15%	65%
RAW 264.7	MAX	75%	45%	55%	<10%	>90%	35%	45%
U2OS	40%	>90%	70%	>90%	MAX	>90%	25%	>90%
KEY	≥80% M/	AX RLUs			>50% to	<80% MA	X RLUs	

KEY

>50% to <80% MAX RLUs

Product	Size	Cat. #
ViaFect™ Transfection Reagent	0.75ml	E4981
	2 x 0.75ml	E4982
FuGENE® HD Transfection Reagent	1ml	E2311
	5 x 1ml	E2312
FuGENE® 6 Transfection Reagent	1ml	E2691
	5 x 1ml	E2692

CellTiter-Fluor, ONE-Glo and ViaFect aretrademarks of Promega Corporation. Products may be covered by issued or pending patents or subject to use limitations, please visit www.promega.com for more information FuGENE is a registered trademark of Fugent, L.L.C.



4