

## **Product Application**

## DNA Extraction From Samples Containing Gelatin Using the ReliaPrep<sup>™</sup> Blood gDNA Miniprep System

DNA was manually purified from samples containing gelatin using the ReliaPrep™ Blood gDNA Miniprep System. Extracted DNA was suitable for speciation applications.

Kit:	ReliaPrep™ Blood gDNA Miniprep System (Cat.# A5081)	This protocol was developed by Promega Applications Scientists and is intended for research use only.
Analyses:	Quantitation by absorbance and with fluorescent dye qPCR amplification	Users are responsible for determining suitability of the protocol for their application.
Sample Type(s):	Aspic, gelatin sheet, capsule and candy	Further information can be found by e-mailing technical services at techserv@promega.com
Input:	Up to 100mg	
Materials Required:	<ul> <li>ReliaPrep<sup>™</sup> Blood gDNA Miniprep System (Cat.# A5)</li> </ul>	081)

- CTAB Buffer (Cat.# MC1411)
- RNase A Solution (Cat.# A7973)
- Proteinase K (PK) solution (Cat.# MC5005)
- Elution Buffer (Cat.# A8281)
- Isopropanol 100%
- Frozen mortar and pestle
- Heat block

## Protocol:

- 1. Grind sample with a frozen mortar and pestle.
- 2. Add 600μl of CTAB Buffer, 2μl of RNase A Solution and 30μl of Proteinase K (PK) Solution to each tube containing up to 100mg of sample. Tap and vigorously vortex the tubes.
- 3. Place tubes in a heat block at 60°C for 30 minutes with shaking at 600rpm. After incubation, vortex tubes with lysate to mix thoroughly.
- 4. Centrifuge the tubes for 10 minutes at  $\geq$ 16,000 × g to separate any solids and oils.
- 5. Transfer 300µl of cleared lysate to a clean 1.5ml microtube.
- 6. Add 300μl of CLD Buffer (Cell Lysis Buffer) to supernatant. Add 600μl of 100% Isopropanol. Vortex to mix.
- Load 600µl of sample to a ReliaPrep<sup>™</sup> Binding Column placed in a collection tube. Centrifuge for 1 minute at maximum speed. Discard flowthrough.
- 8. Load the rest of the sample to the ReliaPrep<sup>™</sup> Binding column and spin for 1 minute more. Place the ReliaPrep<sup>™</sup> Binding Column into a new collection tube.
- 9. Add 500µl of Column Wash Solution (CWD). Spin for 2 minutes at maximum speed. Discard the flowthrough.
- 10. Repeat Step 9 twice for a total of three washes.



 Place the ReliaPrep<sup>™</sup> Binding Column in a labeled elution tube. Add 50µl of Elution Buffer to the ReliaPrep<sup>™</sup> Binding Column. Spin for 1 minute at maximum speed. Discard the column and save eluate.

## **Results:**

Sample	NanoDrop™ ONE (ng/μl)	QuantiFluor <sup>®</sup> ssDNA System (ng/µl)
Aspic	132.38 ± 4.69	71.40 ± 2.61
Gelatin Sheet	7.58 ± 2.63	0.36 ± 0.03
Capsule	22.26 ± 1.83	20.52 ± 1.88
Candy	7.21 ± 0.85	0.63 ± 0.38

Table 1. Concentration and purity ratios of DNA extracted from 100mg of samples containing gelatin using ReliaPrep<sup>™</sup> Blood gDNA Miniprep System (Cat.# A5081). DNA concentration and purity ratios were assessed by absorbance on the NanoDrop<sup>™</sup> One Spectrophotometer and using the QuantiFluor<sup>®</sup> ssDNA System (Cat.# E3190).

Sample	RapidFinder™ Pork ID Kit	RapidFinder™ Beef ID Kit	Expected Origin of Gelatin
Aspic	+	-	Pork
Gelatin Sheet	+	-	Pork
Capsule	-	+	Unknown
Candy	+	_	Unknown
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+ Amplification – No Amplification

Table 2. qPCR amplification of DNA extracted from 100mg of samples containing gelatin using ReliaPrep<sup>™</sup> Blood gDNA Miniprep System (Cat.# A5081). Five microliters of extracted DNA at 10ng/µl (7.58 and 7.21ng/µl for gelatin sheet and candy, respectively) was amplified using RapidFinder<sup>™</sup> Pork ID (Thermo Fisher ref. A24392) and RapidFinder<sup>™</sup> Beef ID (Thermo Fisher ref. A24391).