FuGENE® HD Transfection Reagent

Instructions for Use of Products E2311 AND E2312.

Quick Protocol

Transfection Protocol

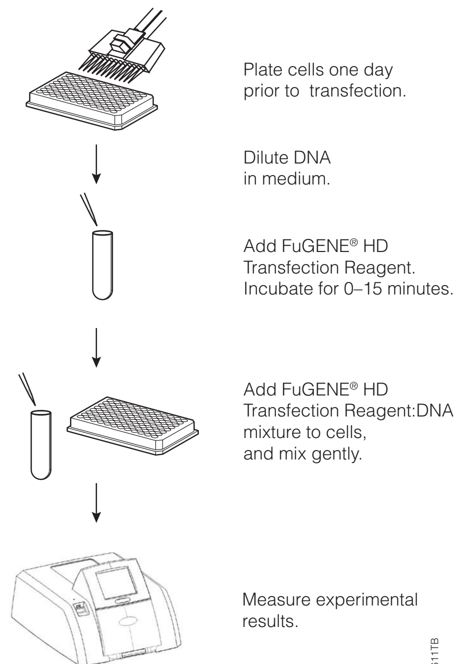
Preparing the FuGENE® HD Transfection Reagent

1. Before use, allow the vial of FuGENE® HD Transfection Reagent to reach room temperature.
2. Mix by inverting or vortexing briefly. If a precipitate is visible, briefly warm at 37°C, then cool to room temperature.

General Transfection Protocol

1. To a sterile tube or U- or V-bottom plate, add 90–98µl of medium prewarmed to room temperature so that the final volume after adding the DNA is 100µl. Add 2µg of plasmid DNA (0.2–1µg/µl) and vortex. For a 3:1 FuGENE® HD Transfection Reagent:DNA ratio, add 6µl of FuGENE® HD Transfection Reagent directly to medium and mix immediately. For other ratios, see the table below. **Note:** Do not allow undiluted FuGENE® HD Transfection Reagent to contact the sides of the tube or U- or V-bottom plate.

	Ratio of FuGENE® HD Transfection Reagent to DNA					
Reagent Volumes or Amounts	4:1	3.5:1	3:1	2.5:1	2:1	1.5:1
Medium to a final volume of	100µl	100µl	100µl	100µl	100µl	100µl
DNA	2µg	2µg	2µg	2µg	2µg	2µg
FuGENE® HD Transfection Reagent	8µl	7µl	6µl	5µl	4µl	3µl



2. Incubate the FuGENE® HD Transfection Reagent/DNA mixture for 0–15 minutes at room temperature.
3. Add 2–10µl of the FuGENE® HD Transfection Reagent/DNA mixture per well to a 96-well plate containing 100µl of cells in growth medium. Mix by pipetting or using a plate shaker. Return cells to the incubator for 24–48 hours.
4. Measure transfection efficiency using an assay appropriate for the reporter gene. For transient transfection, cells are typically assayed 24–48 hours after transfection.

Additional protocol information in Technical Manual #TM328, available online at: **www.promega.com**

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